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PTO/SB/16 (02-01)

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PROVISIONAL APPLICATION FOR PATENT COVER SHEET

This is a request for filing a PROVISIONAL APPLICATION FOR PATENT under 37 CFR 1.53(c).

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60/372450
11 U.S. PTO
04/16/02**INVENTOR(S)**

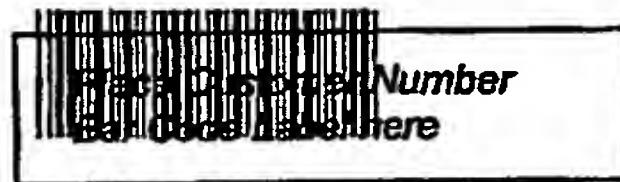
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 Additional Inventors are being named on the 1 separately numbered sheets attached hereto**TITLE OF THE INVENTION (280 characters max)****SYSTEM AND METHOD FOR MOBILE TICKETING OPERATIONS**

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ENCLOSED APPLICATION PARTS (check all that apply) Specification Number of Pages

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 Other (specify) Application Data Sheet. See 37 CFR 1.76**METHOD OF PAYMENT OF FILING FEES FOR THIS PROVISIONAL APPLICATION FOR PATENT** Applicant claims small entity status. See 37 CFR 1.27.**FILING FEE AMOUNT (\$)** A check or money order is enclosed to cover the filing fees

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Respectfully submitted,

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April 15, 2002

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Assistant Commissioner for Patents
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U.S.A.

Dear Sir:

Re: New United States Provisional Patent Application
Title: TB VACCINES
Inventors: LIU, J., CHEN, J., and ALEXANDER, D.C.

We apply in the name of J. Liu, J. Chen, and D.C. Alexander for a provisional patent application entitled **TB VACCINES**.

In addition to the \$150.00 filing fee, included in our firm cheque, we enclose the following documents:

1. provisional application cover page; and
2. patent application

Please direct any questions to Kathryn Schubert at 416-941-9027.

Yours very truly,


Gervas W. Wall
Registration No. 35766

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Encl.

Tuberculosis Vaccines Including Recombinant BCG Strains Expressing Alanine Dehydrogenase, Serine Dehydratase and/or Glutamine Synthetase

Field of the Invention

This invention relates to tuberculosis (TB) vaccines.

Background of the Invention

TB is a deadly contagious disease caused by the infectious agent, *Mycobacterium tuberculosis*. It kills 2 million people each year. The World Health Organization (WHO) 2001 annual report estimated that there would be 8.4 million new TB cases in 1999, up from 8.0 million in 1997. If the present trend continues, it is estimated that between 2000 and 2020, nearly one billion people will be newly infected, 200 million people will become ill and 35 million will die from TB. The spread of HIV/AIDS and the emergence of multidrug-resistant TB contribute to the worsening impact of this disease. Bacille Calmette-Guérin (BCG), an attenuated strain of *Mycobacterium bovis*, is currently the only available vaccine for the prevention of TB. In animal models of infection, BCG vaccination has been demonstrated to induce protective immunity against a *M. tuberculosis* challenge (Baldwins et al., 1998). In humans, BCG vaccination has demonstrated consistent protection against the childhood forms of TB, especially meningitis. However, BCG vaccination is controversial due to variations in its efficacy for protecting adults from pulmonary TB (Fine, 1989; Colditz et al., 1994; Sterne et al., 1998). Trials conducted in the 1940s and 1950s in developed countries such as the United Kingdom, Denmark and North America demonstrated the vaccine to be highly efficient (70-80%). However, in the single largest clinical trial, which took place in India in 1970s and involved more than 265,000 persons, BCG vaccination provided no detectable protection against pulmonary TB. Thus, there is an urgent need to generate an improved vaccine(s) to replace the BCG and to prevent TB.

Several explanations have been suggested for the variation in protective efficacy of BCG (Andersen, 2001). The most prominent hypothesis is that exposure to environmental mycobacteria sensitizes the host against mycobacteria in general, thereby providing

heterologous immunity that obscures the potential benefits of BCG vaccination (Fine, 1995; Fine and Vynnycky, 1998). Furthermore, a recent study showed that the multiplication of BCG was inhibited in animals sensitized with environmental mycobacteria, and consequently BCG vaccination elicited only a transient immune response and failed to provide protective immunity against TB (Brandt et al., 2002). This study also supports the long-standing observation that the induction of immunity to TB requires productive infection by BCG. BCG is a live vaccine; killed BCG does not provide protection. Like *M. tuberculosis*, BCG is capable of forming granulomas and abscesses in various tissues in the infected host (Hogan et al., 2001). The ability of *M. tuberculosis* and *M. bovis* BCG to survive and persist within granulomas, a hostile environment with restricted access to nutrients and reduced oxygen tension, appears to be dependent on the ability of the bacteria to adapt their metabolism to the available source of carbohydrate, nitrogen, and energy (Barclay and Wheeler, 1989). A recent study revealed that fatty acids serve as a source of carbohydrates and are required for persistence of *M. tuberculosis* in mice and activated macrophages (McKinney et al., 2000). Following vaccination in immunocompetent individuals, BCG may persist for certain periods before it is eliminated from the host (Dunn and North 1995; Lagranderie et al., 1996; Moisan et al., 2001).

The key to developing a new and effective TB vaccine is to provide long-term protection (Orme, 2001; Young, 2000). Existing BCG vaccines impart protection against the manifestations of TB in children, but their efficacy wanes over a period of 10 to 15 years, presumably because the protective immunity induced by BCG is gradually lost (Orme, 2001). New strategies to developing an improved vaccine have included the use of attenuated mycobacteria, subunit vaccines and DNA vaccines (Andersen, 2001). However, none of these have proved to be more potent than, or even as effective as BCG. Survival and growth of *M. bovis* BCG is necessary for eliciting protective immunity. It has been shown that early treatment of infected mice with isoniazid to inhibit bacillary growth prevents the development of acquired resistance. BCG strains that persist for extended periods within the host are required in order to obtain more effective vaccines. As such, there is a need for novel, recombinant strains of Bacille Calmette-Guérin.

Summary of the Invention

The invention provides vaccines that overcome the limited ability of BCG strains to use naturally occurring amino acids as the nitrogen source for growth. Furthermore, L-alanine, D-alanine, or L-serine inhibits the growth of BCG strains even when ammonium is present. Expressing a functional alanine dehydrogenase [SEQ ID NO:1; SEQ ID NO: 2] in BCG strains relieves the growth inhibition of BCG by alanine. Similarly, expressing a functional L-serine dehydratase [SEQ ID NO:5; SEQ ID NO: 6] in BCG strains relieves the growth inhibition of BCG by L-serine. The mechanism for such inhibition occurs through blockage of glutamine synthetase. Overexpression of glutamine synthetase [SEQ ID NO:7] to [SEQ ID NO: 14] in BCG relieves the growth inhibition of BCG by alanine and L-serine. Recombinant BCG strains that express (or overexpress) a functional alanine dehydrogenase [SEQ ID NO:1; SEQ ID NO: 2], a L-serine dehydratase [SEQ ID NO:5; SEQ ID NO: 6], and/or glutamine synthetase [SEQ ID NO:7] to [SEQ ID NO: 14] survive and persist longer within the host and consequently induce long-term protective immunity. Such persistent recombinant BCG strains provide more effective vaccines for the prevention of TB and other mycobacterial infections.

The present invention relates to recombinant *Mycobacterium bovis* BCG, which express DNA encoding an alanine dehydrogenase [SEQ ID NO:1; SEQ ID NO: 2], a L-serine dehydratase [SEQ ID NO:5; SEQ ID NO: 6], and/or a glutamine synthetase [SEQ ID NO:7] to [SEQ ID NO: 14]. We found that, due to the lack of a functional alanine dehydrogenase [SEQ ID NO:3; SEQ ID NO: 4], BCG cannot utilize alanine (L-alanine or D-alanine) as the only nitrogen source for growth. We further found that alanine (L-alanine or D-alanine) inhibits the growth of all BCG vaccine strains. Said inhibition is relieved by expressing a functional alanine dehydrogenase [SEQ ID NO:1; SEQ ID NO: 2] in BCG. Similarly, BCG cannot utilize L-serine as the only nitrogen source for growth and that growth of BCG is inhibited by L-serine. Expressing a L-serine dehydratase [SEQ ID NO:5; SEQ ID NO: 6] in BCG strains relieves the growth inhibition by L-serine.

Alanine (L-alanine or D-alanine) and L-serine inhibits BCG growth likely by blocking the activity of glutamine synthetase [SEQ ID NO:7] to [SEQ ID NO: 14]. Overexpression of

glutamine synthetase [SEQ ID NO:7] to [SEQ ID NO: 14] in BCG relieves the growth inhibition of BCG by alanine and L-serine. Glutamine synthetase, in conjunction with glutamate synthase, provides glutamine and glutamate, which are essential for biosynthesis of all amino acids, proteins, purines and pyrimidines. Inhibition of glutamine synthetase stops cell growth. Supplying amino acids that can be converted to glutamate such as L-glutamine, L-glutamate, L-aspartate, and L-asparagine can relieve such inhibition. Indeed, our data show that the inhibition of BCG growth by alanine (L-alanine or D-alanine) or L-serine is relieved by supplementing growth medium with L-glutamine, L-glutamate, L-aspartate, or L-asparagine.

Since BCG is a live vaccine, recombinant BCG strains expressing or overexpressing a functional alanine dehydrogenase [SEQ ID NO:1; SEQ ID NO: 2], a L-serine dehydratase [SEQ ID NO:5; SEQ ID NO: 6], and/or a glutamine synthetase [SEQ ID NO:7] to [SEQ ID NO: 14] survive longer within the human host and subsequently induce long-term memory immunity. These recombinant BCG strains provide extremely useful vaccines.

The present invention relates to a live recombinant *Mycobacterium bovis*-BCG strain comprising a nucleic acid capable of expression, the nucleic acid encoding at least one protein or polypeptide that exhibits alanine dehydrogenase activity [SEQ ID NO:1; SEQ ID NO:2], glutamine synthetase activity [SEQ ID NO:7 to SEQ ID NO:14], or L-serine dehydratase activity [SEQ ID NO:5; SEQ ID NO:6].

The invention also relates to a live recombinant *Mycobacterium bovis*-BCG strain comprising a nucleic acid capable of expression, the nucleic acid encoding at least one protein or polypeptide selected from the group consisting of alanine dehydrogenase [SEQ ID NO:1; SEQ ID NO:2], glutamine synthetase [SEQ ID NO:7 to SEQ ID NO:14] and L-serine dehydratase [SEQ ID NO:5; SEQ ID NO:6].

The invention further relates to a live recombinant *Mycobacterium bovis*-BCG strain comprising a nucleic acid capable of expression, the nucleic acid comprises all or part of at least one nucleic acid molecule selected from the group consisting of [SEQ ID NO:1], [SEQ ID NO:2], [SEQ ID NO:5], [SEQ ID NO:6], [SEQ ID NO:7], [SEQ ID NO:8],

[SEQ ID NO:9], [SEQ ID NO:10], [SEQ ID NO:11], [SEQ ID NO:12], [SEQ ID NO:13] and [SEQ ID NO:14].

In one embodiment, the live recombinant *Mycobacterium bovis*-BCG strain is selected from the group consisting of *Mycobacterium bovis*-BCG-Russia, *Mycobacterium bovis*-BCG-Moreau, *Mycobacterium bovis*-BCG-Japan, *Mycobacterium bovis*-BCG-Sweden, *Mycobacterium bovis*-BCG-Birkhaug, *Mycobacterium bovis*-BCG-Prague, *Mycobacterium bovis*-BCG-Glaxo, *Mycobacterium bovis*-BCG-Denmark, *Mycobacterium bovis*-BCG-Tice, *Mycobacterium bovis*-BCG-Frappier, *Mycobacterium bovis*-BCG-Connaught, *Mycobacterium bovis*-BCG-Phipps, and *Mycobacterium bovis*-BCG-Pasteur.

Another aspect of the invention is a pharmaceutical composition comprising a live recombinant *Mycobacterium bovis*-BCG strain comprising a nucleic acid capable of expression, the nucleic acid encoding at least one protein or polypeptide that exhibits alanine dehydrogenase activity [SEQ ID NO:1; SEQ ID NO:2], glutamine synthetase activity [SEQ ID NO:7 to SEQ ID NO:14], or L-serine dehydratase activity [SEQ ID NO:5; SEQ ID NO:6].

The invention also relates to a live recombinant *Mycobacterium bovis*-BCG strain comprising a nucleic acid capable of expression, the nucleic acid encoding at least one protein or polypeptide selected from the group consisting of alanine dehydrogenase [SEQ ID NO:1; SEQ ID NO:2], glutamine synthetase [SEQ ID NO:7 to SEQ ID NO:14] and L-serine dehydratase [SEQ ID NO:5; SEQ ID NO:6].

In yet another aspect of the invention there is a pharmaceutical composition comprising a live recombinant *Mycobacterium bovis*-BCG strain comprising a nucleic acid capable of expression, the nucleic acid comprises all or part of at least one nucleic acid molecule selected from the group consisting of [SEQ ID NO:1], [SEQ ID NO:2], [SEQ ID NO:5], [SEQ ID NO:6], [SEQ ID NO:7], [SEQ ID NO:8], [SEQ ID NO:9], [SEQ ID NO:10], [SEQ ID NO:11], [SEQ ID NO:12], [SEQ ID NO:13] and [SEQ ID NO:14].

In a futher aspect of the invention there is a vaccine or immunogenic composition for treatment or prophylaxis of a mammal against challenge by mycobacteria comprising a live recombinant *Mycobacterium bovis*-BCG strain comprising a nucleic acid capable of expression, the nucleic acid encoding at least one protein or polypeptide that exhibits alanine dehydrogenase activity [SEQ ID NO:1; SEQ ID NO:2], glutamine synthetase activity [SEQ ID NO:7 to SEQ ID NO:14], or L-serine dehydratase activity [SEQ ID NO:5; SEQ ID NO:6].

In another aspect of the invention there is a vaccine or immunogenic composition for treatment or prophylaxis of a mammal against challenge by mycobacteria comprising a live recombinant *Mycobacterium bovis*-BCG strain comprising a nucleic acid capable of expression, the nucleic acid encoding at least one protein or polypeptide selected from the group consisting of alanine dehydrogenase [SEQ ID NO:1; SEQ ID NO:2], glutamine synthetase [SEQ ID NO:7 to SEQ ID NO:14] and L-serine dehydratase [SEQ ID NO:5; SEQ ID NO:6].

In yet another aspect of the invention there is a vaccine or immunogenic composition for treatment or prophylaxis of a mammal against challenge by mycobacteria comprising a live recombinant *Mycobacterium bovis*-BCG strain comprising a nucleic acid capable of expression, the nucleic acid comprises all or part of at least one nucleic acid molecule selected from the group consisting of [SEQ ID NO:1], [SEQ ID NO:2], [SEQ ID NO:5], [SEQ ID NO:6], [SEQ ID NO:7], [SEQ ID NO:8], [SEQ ID NO:9], [SEQ ID NO:10], [SEQ ID NO:11], [SEQ ID NO:12], [SEQ ID NO:13] and [SEQ ID NO:14]. In a preferred embodiment the vaccine or immunogenic composition is for the treatment or prophylaxis of a mammal against challenge by *Mycobacterium tuberculosis*. In another preferred embodiment the vaccine or immunogenic compositions of the current invention further comprise a pharmaceutically acceptable carrier. In yet another preferred embodiment the vaccine or immunogenic compositions further comprise adjuvants. In a another embodiment the vaccine or immunogenic compositions further comprises immunogenic materials from one or more other pathogens.

Another aspect of this invention relates to a method for treatment or prophylaxis of a mammal against challenge by *Mycobacterium tuberculosis* or *Mycobacterium bovis* comprising administering to the mammal a vaccine or immunogenic composition of the instant invention. In one embodiment the mammal is a cow. In another embodiment the mammal is a human. In yet another embodiment the vaccine or immunogenic composition is administered in the presence of an adjuvant.

A further aspect of the invention is a method for the treatment or prophylaxis of a mammal against cancer comprising administering to the mammal a vaccine or immunogenic composition of the current invention. In one embodiment the cancer is bladder cancer. In another embodiment the vaccine or immunogenic composition is administered in the presence of an adjuvant.

The invention also relates to a test kit comprising the live recombinant *Mycobacterium bovis*-BCG strain of the instant invention.

The invention further relates to a media composition for inhibiting the growth of *Mycobacterium bovis*-BCG comprising alanine as the only nitrogen source for growth. In another embodiment serine is the only nitrogen source for growth. In another embodiment, the media compositions of the current invention further comprise a carbon source, iron, magnesium, and SO₄. In one embodiment the carbon source is selected from the group consisting of glycerol, dextrose, citrate, and glucose.

The current invention relates to a method for inhibiting the growth of *Mycobacterium bovis*-BCG comprising the steps of (a) obtaining a sample comprising *Mycobacterium* and (b) culturing the sample in a selective media. In one embodiment the selective media comprises alanine as the only nitrogen source. In yet another embodiment the selective media comprises serine as the only nitrogen source.

Another aspect of the invention relates to a method for culturing *Mycobacterium bovis*-BCG comprising the steps of (a) obtaining a sample comprising *Mycobacterium* and (b)

culturing the sample in differential media. In one embodiment the differential media comprises histidine.

Brief Description of the Drawings

Preferred embodiments of the invention will be described in relation to the drawings in which:

Fig. 1. Cloning of the *ald* gene. First, a 4.5 kb *Scal*I fragment of *M. tuberculosis* genomic DNA containing the *ald* gene [SEQ ID NO:1] was ligated to *Ecl*136II-linearized pUC19 to generate pUC-ALD. Then, mycobacterial plasmid pALD was created by ligating the 1.9 kb *Kpn*I fragment containing the *ald* gene [SEQ ID NO:1] to *Kpn*I-linearized pMD31.

Fig. 2. Cloning of the *sdaA* gene.

Cloning of *sdaA* [SEQ ID NO:5] was accomplished in two steps. First, a 9.5 kb *Bam*HI fragment of *M. tuberculosis* genomic DNA was ligated to *Bam*HI-linearized pMD31 to generate pSDA1. Plasmid pSDAA was generated by cleavage of pSDA1 with *Pst*I, followed by self-ligation of the 10.9 kb *Pst*I fragment.

Fig. 3. Inhibition of BCG growth by L-alanine in GAS. BCG-Japan, BCG-Frappier, and BCG-Pasteur grown to stationary phase in 7H9/ADC/glycerol/Tween-80 liquid media, were each inoculated into duplicated 5 ml culture volumes of GAS, GAS without L-alanine, and GAS supplemented with 27 mM L-asparagine, to a cell density of 2×10^7 cells/ml. Cultures were incubated at 37°C with constant shaking for 16 days and then 2 ml aliquots of cell culture were centrifuged and cell pellet lyophilized to determine cell dry weight.

Fig. 4. Inhibition of BCG growth by increasing concentrations of L-alanine in Sauton containing NH₄Cl (5 g/liter). a) BCG-Japan, b) BCG-Frappier, and c) BCG-Pasteur, grown to stationary phase in 7H9/ADC/glycerol/Tween-80 liquid media. Cells were washed and resuspended in Sauton basal medium (no nitrogen source).

Resuspended cells of each strain were inoculated into duplicate 5 ml culture volumes of Sauton media supplemented with NH₄Cl and increasing concentrations of L-alanine. Cultures were incubated at 37°C with constant shaking for 30 days and cell dry weight was determined.

Fig. 5. Inhibition of BCG growth by D-alanine in GAS. BCG-Japan, BCG-Frappier, and BCG-Pasteur grown to stationary phase in 7H9/ADC/glycerol/Tween-80 liquid media, were each inoculated into 5ml culture volumes of GAS in which L-alanine was replaced by D-alanine, GAS without L-alanine and, GAS (containing D-alanine) supplemented with 27 mM L-asparagine, to a cell density of 2×10^7 cells/ml. Cultures were incubated at 37°C with constant shaking for 13 days and cell dry weight was determined.

Fig. 6. Growth of recombinant BCG strains expressing alanine dehydrogenase [SEQ ID NO:1] in GAS medium. The growth of BCG-Frappier/*ald*, BCG-Pasteur/*ald*, BCG-Frappier/pMD31, BCG-Pasteur/pMD31, BCG-Frappier, and BCG-Pasteur were compared. Cells of each strain, grown to stationary phase in 7H9/ADC/glycerol/Tween-80 liquid media, were washed and resuspended in Sauton basal medium (no nitrogen source). Resuspended cells were inoculated into duplicate 5 ml culture volumes of GAS without L-alanine, GAS containing L-alanine and GAS in which L-alanine was replaced by D-alanine. Cultures were incubated at 37°C with constant shaking for 15 days and cell dry weight was then determined.

Fig. 7. Inhibition of BCG growth by L-serine in GAS. BCG-Japan, BCG-Frappier, and BCG-Pasteur grown to stationary phase in 7H9/ADC/glycerol/Tween-80 liquid media, were each inoculated into duplicate 5 ml culture volumes of GAS in which L-alanine was replaced by L-serine, GAS without L-alanine, and GAS (containing L-serine) supplemented with 27 mM L-asparagine, to a cell density of 2×10^7 cells/ml. Cultures were incubated at 37°C with constant shaking for 15 days and cell dry weight was then determined.

Fig. 8. Growth of recombinant BCG strains expressing L-serine dehydratase [SEQ ID NO:5] in GAS medium containing L-serine. The growth of BCG-Japan/*sdaA*,

BCG-Frappier/*sdaA*, BCG-Pasteur/*sdaA*, BCG-Japan, BCG-Frappier, and BCG-Pasteur were compared. Cells of each strain, grown to stationary phase in 7H9/ADC/glycerol/Tween-80 liquid media, were washed and resuspended in Sauton basal medium (no nitrogen source). Resuspended cells were inoculated into duplicate 5 ml culture volumes of GAS without L-alanine, GAS in which L-alanine was replaced by L-serine, and GAS (containing L-serine) supplemented with 27 mM L-asparagine. Cultures were incubated at 37°C with constant shaking for 15 days and cell dry weight was then determined.

Fig. 9. Alignment of A) nucleotide and B) amino acid sequences of the *ald* genes of *Mycobacterium tuberculosis* (*M. tb*) [SEQ ID NO:1; SEQ ID NO:2] and *Mycobacterium bovis* (*M. bovis*) [SEQ ID NO:3; SEQ ID NO:4] . The point deletion causing the frameshift mutation in *M. bovis* *ald* [SEQ ID NO:3] is indicated with an arrow. Nucleotide codons and amino acids affected by this mutation are highlighted.

Detailed Description of the Invention

BCG vaccine strains have a limited ability to utilize amino acids as the nitrogen source for growth. Furthermore, we found that naturally occurring amino acids L-alanine and L-serine inhibit the growth of BCG strains. Expressing a functional L-alanine dehydrogenase [SEQ ID NO:1; SEQ ID NO:2] in BCG relieves the growth inhibition by alanine. Expressing of a functional L-serine dehydratase [SEQ ID NO:5; SEQ ID NO:6] in BCG relieves the growth inhibition by L-serine. As well, overproduction of glutamine synthetase [SEQ ID NO:7] to [SEQ ID NO: 14] relieves the growth inhibition by alanine and serine. These novel findings are significant because recombinant BCG strains that express (or overexpress) a functional alanine dehydrogenase [SEQ ID NO:1; SEQ ID NO:2], a L-serine dehydratase [SEQ ID NO:5; SEQ ID NO:6] , and/or glutamine synthetase [SEQ ID NO:7] to [SEQ ID NO: 14] will survive better within the human host, induce long-term memory immunity and provide for more effective vaccines to prevent TB, particularly for protecting against pulmonary TB in adults.

It has long been known that administration of killed BCG strains results in a weak and transient immune response. Protective immunity requires survival and replication of BCG

in the vaccinated host. This notion is reinforced by a recent study of an animal model of infection, which showed that prior exposure to live environmental mycobacteria blocked the multiplication of BCG in infected mice. Consequently BCG elicited only a transient immune response which failed to provide protective immunity against TB (Brandt et al., 2002). Live BCG continuously secrete many different antigens that are likely important for the induction of protective immunity. The continuous production of numerous antigens by multiplying BCG gives live vaccines an advantage over subunit vaccines or DNA vaccines which transiently produce a few antigens. Thus the ability of BCG to multiply and persist within the host is an important determinant of BCG efficacy.

In order to grow and persist within the host, BCG must be able to utilize the available nutrients inside the host. It was demonstrated that isocitrate lyase, an essential enzyme for catabolism of fatty acids, is required for persistence of *M. tuberculosis* during the chronic phase of infection and that this requirement was dependent on an intact immune response of the host (McKinney et al., 2000). In another study, an *M. bovis* BCG strain lacking anaerobic nitrate reductase, an enzyme essential for nitrate respiration, failed to persist in lungs, liver and kidneys of immune-competent mice (Fritz et al., 2002). Our findings, that BCG strains utilize only a few types of amino acids as the nitrogen source for growth, and that the growth of all BCG strains are inhibited by naturally occurring L-alanine and L-serine, suggest that the ability of BCG to grow and persist within the host is restricted. The concentration of L-alanine that is available to BCG growing in human is estimated to be 0.33-0.42 mM (Barclay and Wheeler, 1989), which is sufficient to inhibit the growth of BCG-Pasteur or BCG-Frappier, and significantly reduce the growth of BCG-Japan (Fig. 4). The concentration of L-serine present in the extracellular fluids of the host is around 0.1 mM (Barclay and Wheeler, 1989), which may cause significant inhibition of BCG growth. Since multiplication of BCG is required to generate protective immunity, such inhibition by amino acids within the host may prevent the development of long-term protective immunity and hence the lack of protection against pulmonary TB in adults.

M. bovis BCG is also used in the treatment of bladder cancer. Numerous randomized controlled clinical trials indicate that intravesical administration of BCG can prevent or delay tumour recurrence (reviewed in Lamm, 2000; Lockyer and Gillatt, 2001). The

details of how BCG exerts this effect remain to be determined. However, the antitumour response requires an intact T-cell response, and involves increased expression of Th1-type cytokines, including TNF α and IL-6 (reviewed in Prescott et al, 2000). The most effective treatment regimes involve multiple applications of BCG, which suggests that prolonged exposure to the bacteria is required. Similarly, tumours that retain the ability to phagocytize BCG are most susceptible to this treatment (de Boer et al 1996), indicating that bacterial interactions with the tumour are important. As such, a BCG strain demonstrating increased persistence may provide enhanced antitumour activity.

We show that the absence of a functional alanine dehydrogenase [SEQ ID NO:1; SEQ ID NO:2] is responsible for the failure of BCG strains to utilize alanine (L-alanine or D-alanine) as the only nitrogen source. A gene (Rv2708) coding for a L-alanine dehydrogenase (*ald*) [SEQ ID NO:1] was identified in the genome of *M. tuberculosis*. The activity of this enzyme from *M. tuberculosis* had been demonstrated biochemically *in vitro*. Ald converts L-alanine to pyruvate and ammonium, and is highly specific for L-alanine (Hutter and Singh, 1999). This enzyme was detected in the culture supernatant fraction of *M. tuberculosis* but not in *M. bovis* BCG-Japan nor BCG-Copenhagen, even though DNA Southern blot showed that the gene is present in both BCG strains (Anderson et al., 1992). Similarly, we do not detect alanine dehydrogenase activity in any of the 12 BCG strains listed in this report (data not shown). This lack of a functional alanine dehydrogenase in BCG strains is probably caused by a mutation within the *ald* gene [SEQ ID NO:3], and probably originated with the original *M. bovis* strain. A frame-shift mutation is found within the *ald* gene in the published genome sequence of *M. bovis* (Fig. 9) [SEQ ID NO:3]. As a result, the full length L-alanine dehydrogenase protein [SEQ ID NO:2; SEQ ID NO:4] cannot be made in BCG strains and subsequently BCG cannot catabolize alanine. Similarly, the failure of BCG to utilize L-serine as the only nitrogen source is likely to be caused by either mutations or altered expression of the *sdaA* gene [SEQ ID NO:5; SEQ ID NO:6], which encodes L-serine dehydratase. Expression of *sdaA* [SEQ ID NO:5; SEQ ID NO:6] of *M. tuberculosis* in BCG allows BCG strains to grow on L-serine as the only nitrogen source and relieves the inhibition of BCG growth by L-serine (Fig. 8). The inhibition of BCG growth by alanine and serine is

caused by inhibition of glutamine synthetase [SEQ ID NO:7] to [SEQ ID NO: 14]. Overexpression of a glutamine synthetase [SEQ ID NO:7] to [SEQ ID NO: 14] in BCG relieves the growth inhibition by L-serine, L-alanine and D-alanine.

BCG-Frappier and BCG-Pasteur are more susceptible than BCG-Japan to inhibition by alanine, presumably due to difference in the expression level or activity of glutamine synthetase. BCG-Japan differs from BCG-Frappier or BCG-Pasteur genetically (Behr et al., 1999). Calmette and Guérin developed the BCG vaccine in 1921 after 13 years and 230 passages of an isolate of *M. bovis* *in vitro*. Starting from 1924, BCG lots were distributed to laboratories around the world. These laboratories continued the passage of the bacteria *in vitro* employing a variety of different recipes and protocols until 1961 when lyophilized seeds were established. As a consequence of such practices, different BCG progeny strains were created, which differed biochemically and genetically (Oettinger et al., 1999; Behr et al., 1999). Our data show that the ability of BCG strains to utilize amino acids as nitrogen source vary; for example, BCG-Japan is able to grow on cationic amino acids including L-arginine and L-lysine while BCG-Pasteur and BCG-Frappier cannot. These differences may also contribute to the differences of BCG efficacy in various clinical trials.

In summary, we use recombinant BCG strains that express (or overexpress) a functional alanine dehydrogenase [SEQ ID NO:1; SEQ ID NO:2], a L-serine dehydratase [SEQ ID NO:5; SEQ ID NO:6], and/or glutamine synthetase [SEQ ID NO:7] to [SEQ ID NO: 14] as vaccines to prevent TB and other mycobacterial infections. These recombinant BCG vaccines will induce long-term protective immunity against TB.

Variations of Nucleic Acid Molecules

Modifications

Many modifications may be made to the nucleic acid molecule DNA sequences disclosed in this application and these will be apparent to one skilled in the art. The invention includes nucleotide modifications of the sequences disclosed in this application (or fragments thereof) that are capable of directing expression in bacterial or mammalian

cells. Modifications include substitution, insertion or deletion of nucleotides or altering the relative positions or order of nucleotides.

Sequence Identity

The nucleic acid molecules of the invention also include nucleic acid molecules (or a fragment thereof) having at least about: 70% identity, at least 80% identity, at least 90% identity, at least 95% identity, at least 96% identity, at least 97% identity, at least 98% identity or, most preferred, at least 99% or 99.5% identity to a nucleic acid molecule of the invention and which are capable of expression of nucleic acid molecules in bacterial or mammalian cells. Identity refers to the similarity of two nucleotide sequences that are aligned so that the highest order match is obtained. Identity is calculated according to methods known in the art. For example, if a nucleotide sequence (called "Sequence A") has 90% identity to a portion of [SEQ ID NO: 1], then Sequence A will be identical to the referenced portion of [SEQ ID NO: 1] except that Sequence A may include up to 10 point mutations (such as substitutions with other nucleotides) per each 100 nucleotides of the referenced portion of [SEQ ID NO: 1].

Sequence identity (each construct preferably without a coding nucleic acid molecule insert) is preferably set at least about: 70% identity, at least 80% identity, at least 90% identity, at least 95% identity, at least 96% identity, at least 97% identity, at least 98% identity or, most preferred, at least 99% or 99.5% identity to the sequences provided in SEQ ID NO:1 to SEQ ID NO:14 or its complementary sequence). Sequence identity will preferably be calculated with the GCG program from Bioinformatics (University of Wisconsin). Other programs are also available to calculate sequence identity, such as the Clustal W program (preferably using default parameters; Thompson, JD et al., Nucleic Acid Res. 22:4673-4680), BLAST P, BLAST X algorithms.

Hybridization

The invention includes DNA that has a sequence with sufficient identity to a nucleic acid molecule described in this application to hybridize under stringent hybridization conditions (hybridization techniques are well known in the art). The present invention

also includes nucleic acid molecules that hybridize to one or more of the sequences in [SEQ ID NO:1] to [SEQ ID NO:14] or its complementary sequence. Such nucleic acid molecules preferably hybridize under high stringency conditions (see Sambrook et al. *Molecular Cloning: A Laboratory Manual*, Most Recent Edition, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y.). High stringency washes have preferably have low salt (preferably about 0.2% SSC) and a temperature of about 50-65 °C.

Vaccines

One skilled in the art knows the preparation of live recombinant vaccines. Typically, such vaccines are prepared as injectables, either as liquid solutions or suspensions; solid forms suitable for solution in, or suspension in, liquid prior to injection may also be prepared. The preparation may also be emulsified, or the protein encapsulated in liposomes. The live immunogenic ingredients are often mixed with excipients that are pharmaceutically acceptable and compatible with the active ingredient. Suitable excipients are, for example, water, saline, dextrose, glycerol, ethanol, or the like and combinations thereof. In addition, if desired, the vaccine may contain minor amounts of auxiliary substances such as wetting or emulsifying agents, pH buffering agents, and/or adjuvants that enhance the effectiveness of the vaccine. Examples of adjuvants which may be effective include but are not limited to: aluminum hydroxide, N-acetyl-muramyl-L-threonyl-D-isoglutamine (thr-MDP), N-acetyl-nor-muramyl-L-alanyl-D-isoglutamine (CGP 11637, referred to as nor-MDP), N-acetylmuramyl-L-alanyl-D-isoglutaminyl-L-alanine-2-(1'-2'-dipalmitoyl-sn-glycero-3-hydroxyphosphoryloxy)-ethylamine (CGP 19835A, referred to as MTP-PE), and RIBI, which contains three components extracted from bacteria, monophosphoryl lipid A, trehalose dimycolate and cell wall skeleton (MPL+TDM+CWS) in a 2% squalene/Tween 80™ emulsion.

The effectiveness of an adjuvant may be determined by measuring the amount of antibodies directed against an immunogenic polypeptide containing a *Mycobacterium tuberculosis* antigenic sequence resulting from administration of the live recombinant *Mycobacterium bovis*-BCG vaccines that are also comprised of the various adjuvants. The vaccines are conventionally administered parenterally, by injection, for example, either subcutaneously or intramuscularly. Additional formulations which are suitable for

other modes of administration include suppositories and, in some cases, oral formulations. For suppositories, traditional binders and carriers may include, for example, polyalkylene glycols or triglycerides; such suppositories may be formed from mixtures containing the active ingredient in the range of 0.5% to 10%, preferably 1%-2%. Oral formulations include such normally employed excipients as, for example, pharmaceutical grades of mannitol, lactose, starch, magnesium stearate, sodium saccharine, cellulose, magnesium carbonate, and the like. These compositions take the form of solutions, suspensions, tablets, pills, capsules, sustained release formulations or powders and contain 10%-95% of active ingredient, preferably 25%-70%.

The vaccines are administered in a manner compatible with the dosage formulation, and in such amount as will be prophylactically and/or therapeutically effective.

The vaccine may be given in a single dose schedule, or preferably in a multiple dose schedule. A multiple dose schedule is one in which a primary course of vaccination may be with 1-10 separate doses, followed by other doses given at subsequent time intervals required to maintain and or reinforce the immune response, for example, at 1-4 months for a second dose, and if needed, a subsequent dose(s) after several months. The dosage regimen will also, at least in part, be determined by the need of the individual and be dependent upon the judgment of the practitioner.

In addition, the live recombinant *Mycobacterium bovis*-BCG vaccine administered in conjunction with other immunoregulatory agents, for example, immune globulins. A subject of the present invention is also a multivalent vaccine formula comprising, as a mixture or to be mixed, a live recombinant *Mycobacterium bovis*-BCG vaccine as defined above with another vaccine, and in particular another recombinant live recombinant *Mycobacterium bovis*-BCG vaccine as defined above, these vaccines comprising different inserted sequences.

Pharmaceutical compositions

The pharmaceutical compositions of this invention are used for the treatment or prophylaxis of a mammal against challenge by *Mycobacterium tuberculosis* or *Mycobacterium bovis*. The pharmaceutical compositions of this invention are also used to treat patients having degenerative diseases, disorders or abnormal physical states such as cancer.

The pharmaceutical compositions can be administered to humans or animals by methods such as tablets, aerosol administration, intratracheal instillation and intravenous injection.

Media Compositions

The media compositions of this invention for inhibiting the growth of *Mycobacterium bovis*-BCG comprise alanine or serine as the only nitrogen source. When alanine is the only nitrogen source it is present in an amount of at least 0.03mM and when serine is the only nitrogen source it is present in an amount of at least 0.03mM.

The media compositions may further contain carbon in an amount of about 1.35g/L to about 1.65g/L, preferably in an amount of at least 1.5g/L; iron in an amount of about 0.045g/L to about 0.055g/L, preferably in an amount of at least 0.05g/L; magnesium in an amount of about 0.45g/L to about 0.55g/L, prefereably in an amount of at least 0.5g/L; and SO₄ in an amount of about 0.045g/L to about 0.055g/L, prefereably in an amount of at least 0.05g/L.

Kits

Kits suitable for immunodiagnosis and containing the appropriate labeled reagents are constructed by packaging the appropriate materials, including the live recombinant *Mycobacterium bovis*-BCG strains of the instant invention, in suitable containers, along with the remaining reagents and materials required for the conduct of the assay, as well as a suitable set of assay instructions. Any immunological test format is contemplated, such as ELISA, Western blot, sandwich assay etc., which are well known to those skilled in the art.

Materials and Methods

Bacterial strains and culture conditions. Twelve *M. bovis* BCG strains: BCG-Japan, BCG-Russia, BCG-Moreau, BCG-Sweden, BCG-Birkhaug BCG-Frappier, BCG-Pasteur, BCG-Glaxo, BCG-Phipps, BCG-Tice, BCG-Denmark, and BCG-Prague were used in this study and were obtained from Dr. Marcel Behr (McGill University). The identities of these strains were described in detail previously (Behr et al., 1999). Middlebrook 7H9 medium (Difco) contains (per liter) ammonium sulfate, 0.5 g; L-glutamate, 0.5 g; sodium citrate 0.1 g; pyridoxine, 1 mg; biotin, 0.5 mg; disodium phosphate 2.5g; monopotassium phosphate, 1 g; ferric ammonium citrate 40 mg; magnesium sulfate 50 mg; calcium chloride 0.5 mg; zinc sulfate 1 mg; copper sulfate, 1 mg; and glycerol, 2 ml; with 5 g of albumin (fraction V; bovine), 2 g of dextrose, and 0.05% Tween 80 added after sterilization. Sauton medium contains (per liter) L-asparagine, 4 g; monopotassium sulfate, 0.5 g; magnesium sulfate 0.5 g; ferric ammonium citrate 50 mg; citric acid, 2 g; zinc sulfate, 1 mg; and glycerol, 60 ml; with 0.05% Tween 80 added after sterilization. Glycerol-alanine-salts (GAS) medium contains (per liter) 2 g of ammonium chloride, 1 g of L-alanine, 0.3 g of Bacto Casitone (Difco), 4 g of dibasic potassium phosphate, 2 g of citric acid, 50 mg of ferric ammonium citrate, 1.2 g of magnesium chloride hexahydrate, 0.6 g of potassium sulfate, 1.8 ml of 10 M sodium hydroxide, and 10 ml of glycerol. Tween 80 was added to 0.05% after sterilization. BCG cultures were grown at 37°C with constant shaking for 3-4 weeks.

Cloning of *ald*. Cloning of *ald* [SEQ ID NO:1] was accomplished in two steps (Fig. 1). First, a 4.5kb *Scal*I fragment of *M. tuberculosis* genomic DNA containing *ald* was ligated to *Ecl*136II-linearized pUC19 to generate pUC-ALD. Then mycobacterial plasmid pALD was created by ligating the 1.9 kb *Kpn*I fragment containing the *ald* gene [SEQ ID NO:1] to *Kpn*I- linearized pMD31 (Yu et al., 1998). The plasmid pALD was introduced by electroporation into *M. bovis* BCG, and recombinant *M. bovis* BCG selected on Middlebrook 7H9 agar (Difco) supplemented with 10% oleic/albumin/dextrose/catalase (OADC) enrichment and 25 µg/ml kanamycin.

Cloning of *sdaA*. Cloning of *sdaA* [SEQ ID NO:5] was accomplished in two steps. First, a 9.5 kb *Bam*HI fragment of *M. tuberculosis* genomic DNA was ligated to *Bam*HI-linearized pMD31 to generate pSDA1. Plasmid pSDAA was generated by cleavage of pSDA1 with *Pst*I, followed by self-ligation of the 10.9 kb *Pst*I fragment. The plasmid pSDAA was introduced by electroporation into *M. bovis* BCG, and recombinant *M. bovis* BCG selected on Middlebrook 7H9 agar (Difco) supplemented with 10% oleic/albumin/dextrose/catalase (OADC) enrichment and 25 µg/ml kanamycin.

Example 1

Growth of BCG strains in Glycerol-Alanine-Salts (GAS) medium. During the course of our studies, we found that BCG-Japan strain was able to grow in GAS medium, albeit slower than in 7H9 medium. BCG-Frappier and BCG-Pasteur could not grow in GAS medium, even after prolonged incubation (2 months). The growth of other BCG strains in GAS medium was also examined. The results are summarized in Table I, and show that BCG-Japan, BCG-Russia, BCG-Moreau, BCG-Sweden and BCG-Birkhaug were able to grow in GAS medium while BCG-Frappier, BCG-Pasteur, BCG-Glaxo, BCG-Phipps, BCG-Tice, BCG-Denmark, and BCG-Prague could not. This is an interesting observation since all 12 BCG strains listed above were able to grow in 7H9 and Sauton broth medium (Table I). To find out why certain BCG strains were unable to grow in GAS medium, the chemical compositions of GAS, 7H9 and Sauton medium were compared. Supplementing ZnSO₄ (1 mg/liter), which is present in 7H9 and Sauton but not in GAS medium, or sodium pyruvate (0.5%), which is required for growth of large colonies of *M. bovis*, did not support the growth of BCG strains in GAS (data not shown). Next, nitrogen sources were compared. L-Asparagine (4 g/liter) is the only nitrogen source in Sauton medium while ammonium chloride (2 g/liter) and L-alanine (1 g/liter) are the main nitrogen sources in GAS. When L-asparagine (at 4 g per liter) was added to GAS medium, BCG-Frappier, BCG-Pasteur, BCG-Glaxo, BCG-Phipps, BCG-Tice, BCG-Denmark, and BCG-Prague were able to grow rapidly (Table I). Supplementing L-aspartate, L-glutamine, or L-glutamate but not other types of amino acids to GAS medium also supported the growth of these BCG strains (Table I). These results show that the failure

of certain BCG strains to grow in GAS medium is caused by their inability to utilize the nitrogen source present.

Example 2

Amino acids as the nitrogen source for growth of BCG strains. The above result prompted us to examine the ability of BCG strains to utilize various types of amino acids as the only nitrogen source. Since GAS medium contains a small amount of Bacto Casitone (0.3 g/liter), which is a complex mixture of various amino acids and peptides, we chose Sauton medium, which is a defined medium, for this purpose. The L-asparagine in the original formula for Sauton medium was replaced individually by each type of amino acids at the same concentration (27 mM), and pH was adjusted to 7.0. Ammonium chloride at 27 mM or 1 mM as the only nitrogen source was also tested. Table II summarizes the results for three representative BCG strains, BCG-Japan, BCG-Pasteur, and BCG-Frappier. Consistent with the result in Table I, all three BCG strains grew rapidly when L-asparagine, L-aspartate, L-glutamine, or L-glutamate was used as the only nitrogen source. BCG-Japan was able to grow on cationic amino acids (e.g., L-arginine, L-lysine) while BCG-Pasteur and BCG-Frappier could not. More interestingly, none of the BCG strains were able to utilize L-alanine, L-serine, L-leucine, L-isoleucine, L-methioine, or L-glycine as the only nitrogen source, while other *Mycobacterium* species, including pathogenic *M. tuberculosis* and *M. avium*, and nonpathogenic *M. smegmatis*, were able grow on these amino acids. These results demonstrate that BCG vaccine strains utilize limited types of amino acids as the nitrogen source for growth; some BCG strains such as BCG-Pasteur or BCG-Frappier can grow only on 4 types of amino acids (Table II). Such a limitation is likely to restrict the ability of BCG to grow and persist *in vivo* (within the host).

Example 3

L-Alanine, D-alanine, or L-serine inhibits the growth of BCG. One surprising finding from the above experiment was that all BCG strains are able to grow on ammonium chloride as the only nitrogen source at both low (1 mM) or high concentrations (27 mM) (Table II). This is contradictory to the result obtained in GAS medium, in which

ammonium chloride at 37 mM does not support the growth of BCG-Pasteur and BCG-Frappier (Table I). Since GAS medium also contains L-alanine, and L-alanine is not utilized by BCG strains for growth (Table II), the only possible explanation is that L-alanine actually inhibits the growth of BCG strains. To prove this, a modified GAS medium, in which L-alanine was omitted, was made and the growth of BCG strains in this medium was examined. As predicted, BCG-Frappier and BCG-Pasteur, which are unable to grow in the original GAS medium containing L-alanine, grew rapidly in GAS without L-alanine (Fig. 3). BCG-Japan also grew more rapidly in this L-alanine free medium than in the original GAS medium (Fig. 3). The same results were obtained for the other nine BCG strains listed in this report.

To further confirm this result, increasing concentrations of L-alanine were added to Sauton medium containing ammonium chloride (5 g/liter) and the growth of BCG-Japan, BCG-Frappier and BCG-Pasteur was determined (Fig. 4). Strikingly, even at a very low concentration (0.25 mM), L-alanine completely inhibited the growth of BCG-Frappier and BCG-Pasteur. Although the growth inhibition of BCG-Japan was somewhat less severe, L-alanine at 0.5 mM significantly reduced its growth and at 8-16 mM the growth was completely inhibited (Fig. 4). Taken together, these results clearly demonstrate that L-alanine inhibits the growth of BCG strains. We further found that D-alanine also inhibits the growth of BCG strains. The presence of D-alanine in GAS medium stopped the growth of BCG-Pasteur and BCG-Frappier, and significantly reduced the growth of BCG-Japan (Fig. 5). Similarly, the presence of L-serine in GAS medium significantly inhibited the growth of BCG-Japan, BCG-Frappier, and BCG-Pasteur (Fig. 7).

Example 4

Expressing L-alanine dehydrogenase [SEQ ID NO:1; SEQ ID NO:2] in BCG relieves the inhibition of BCG growth by L-alanine and D-alanine. Alanine is an excellent source of nitrogen for many *Mycobacterium* species including *M. tuberculosis*, *M. avium*, and *M. smegmatis*. D-Alanine degradation begins with racemization to L-alanine, which is then broken down to ammonium and pyruvate by L-alanine dehydrogenase. Interestingly, a functional L-alanine dehydrogenase was detected in *M.*

tuberculosis and *M. smegmatis* but not in BCG-Japan or BCG-Copenhagen (Andersen et al., 1992; Hutter and Dick, 1998). We did not detect L-alanine dehydrogenase activity in any of the BCG strains listed in this study (data not shown). The failure of BCG strains to utilize L- or D- alanine as the only nitrogen source for growth is due to the lack of a functional L-alanine dehydrogenase. To prove this, the *ald* gene [SEQ ID NO:1] coding for L-alanine dehydrogenase [SEQ ID NO:2] in the *M. tuberculosis* genome was cloned into a shuttle vector and transformed into BCG-Frappier and BCG-Pasteur. The resulting recombinant BCG strains were tested for their ability to grow in GAS medium containing L-alanine or D-alanine. Both recombinant strains, BCG-Frappier/*ald* and BCG-Pasteur/*ald*, grew rapidly in GAS medium containing either L-alanine or D-alanine (Fig. 6), while strains containing the cloning vector alone did not grow. This result shows that expression of a functional L-alanine dehydrogenase [SEQ ID NO:1; SEQ ID NO:2] in BCG strains relieves the growth inhibition of BCG by L-alanine and D-alanine.

Example 5

Expressing L-serine dehydratase [SEQ ID NO:5; SEQ ID NO:6] in BCG relieves the inhibition of BCG growth by L-serine. L-Serine is used by *M. tuberculosis*, *M. avium* and *M. smegmatis*, but not *M. bovis* BCG, as the only nitrogen for growth. The failure of BCG to utilize L-serine as the only nitrogen source is likely to be caused by either mutations on or altered expression of the gene encoding L-serine dehydratase, *sdaA* [SEQ ID NO:5], in BCG. Expression of *sdaA* [SEQ ID NO:5; SEQ ID NO:6] of *M. tuberculosis* in BCG allows BCG strains to grow on L-serine as the only nitrogen source and relieves the inhibition of BCG growth by L-serine (Fig. 8).

Example 6

Inhibition of BCG growth by L-alanine, D-alanine and L-serine are likely to occur by blocking the activity of glutamine synthetase [SEQ ID NO:7] to [SEQ ID NO:14]. Glutamine synthetase plays a central role in nitrogen metabolism in bacteria (Reitzer, 1996). Working in tandem with glutamate synthase, glutamine synthetase catalyzes the synthesis of glutamine and glutamate, which together provide nitrogen for almost all amino acids, proteins, and nucleotides. In *Escherichia coli* and *Klebsiella aerogenes*,

glutamine synthetase is under feedback inhibition – purified glutamine synthetase is inhibited by L-alanine, L-serine and glycine (Reitzer, 1996). Glutamine synthetase was identified as an extracellular protein in *M. tuberculosis* and *M. bovis* BCG (Harth et al., 1994). It is likely that undegraded L-alanine inhibits glutamine synthetase and subsequently prevents the growth of BCG. If this were correct, then L-serine, which was not catabolized by BCG for growth (Table I), would also inhibit the growth of BCG by the same mechanism. Supporting this hypothesis, addition of L-serine to GAS medium containing only ammonium chloride as the nitrogen source inhibits the growth of BCG-Frappier, BCG-Pasteur or BCG-Japan (Fig. 7). Furthermore, if glutamine synthetase were the target of L-alanine and L-serine inhibition, then supplementing amino acids that can be converted to glutamate would also alleviate their effects, as demonstrated in *K. aerogenes* (Janes and Bender, 1998). Indeed, addition of L-glutamate and amino acids that could be catabolized to yield glutamate (L-glutamine, L-asparagine, and L-aspartate) allows the growth of BCG strains in the presence of alanine (Table I), but those that could not be catabolized to glutamate (e.g., L-lysine, L-methioine, L-leucine) fail to allow growth. BCG-Frappier and BCG-Pasteur are more sensitive than BCG-Japan to inhibition by alanine and serine, this is due to differences in the expression level or activity of glutamine synthetase [SEQ ID NO:7] to [SEQ ID NO:14], i.e., BCG-Japan produces more glutamine synthetase or with higher activity than BCG-Frappier or BCG-Pasteur.

The present invention has been described in detail and with particular reference to the preferred embodiments; however, it will be understood by one having ordinary skill in the art that changes can be made without departing from the spirit and scope thereof. For example, where the application refers to proteins, it is clear that peptides and polypeptides may often be used. Likewise, where a gene is described in the application, it is clear that nucleic acids or gene fragments may often be used.

All publications (including Genbank entries), patents and patent applications are incorporated by reference in their entirety to the same extent as if each individual publication, patent or patent application was specifically and individually indicated to be incorporated by reference in its entirety.

Table I

Comparative growth of *M. tuberculosis*, *M. smegmatis* and *M. bovis* BCG substrains in 7H9, Sauton, and glycerol-alanine-salts (GAS) medium.

Mycobacterium ^a	7H9	Sauton	GAS	GAS + L-Asn ^b	GAS + L-Asp ^b	GAS + L-Gln ^b
<i>M. tuberculosis</i> ^c	+	+	+	+	+	+
<i>M. smegmatis</i>	+	+	+	+	+	+
BCG-Russia	+	+	+	+	+	+
BCG-Moreau	+	+	+	+	+	+
BCG-Japan	+	+	+	+	+	+
BCG-Sweden	+	+	+	+	+	+
BCG-Birkhaug	+	+	-	-	-	-
BCG-Prague	+	+	+	+	+	+
BCG-Glaxo	+	-	-	-	-	-
BCG-Denmark	+	-	-	-	-	-

BCG-Tice	+	+	-	-	+	+	+	+
BCG-Frappier	+	+	-	-	+	+	+	+
BCG-Phipps	+	+	-	-	+	+	+	+
BCG-Pasteur	+	+	-	-	+	+	+	+

^a Each 5 ml culture inoculated with 1×10^7 cells of *M. smegmatis* or *M. bovis* BCG substrains.

^b L-Asn, L-Asp, L-Glu and L-Gln in GAS supplemented to a final concentration of 27 mM.

^c Based on research literature.

Table II

Comparative growth of *M. bovis* BCG-Japan, BCG-Frappier, BCG-Pasteur, *M. tuberculosis*, *M. avium* and *M. smegmatis*

Media ^a	BCG-Japan ^b	BCG-Frappier ^b	BCG-Pasteur ^b	<i>M. tuberculosis</i> ^c	<i>M. avium</i> ^c	<i>M. smegmatis</i> ^b
Sauton basal						
Group 1						
Sauton + L-Asn	+++	+++	+++	+++	+++	+++
Sauton + L-Asp	+++	+++	+++	+++	+++	+++
Sauton + L-Glu	+++	+++	+++	+++	+++	+++
Sauton + L-Gln	+++	+++	+++	+++	+++	+++
Sauton + L-Cys	+++	+++	+++	+++	+++	+++
Sauton + NH ₄ Cl	++	++	++	++	++	++
Group 2						
Sauton + L-Arg	++					+++
Sauton + L-His	++					+++

Sauton + L-Lys	++	NA	+++	+++
Sauton + L-Pro	++	NA	-	++
Sauton + GABA	++	NA	NA	+++
Sauton + L-Ornithine	++	NA	NA	+++
Group 3				
Sauton + L-Ala	+++	+++	+++	+++
Sauton + L-Ser	++	++	++	++
Sauton + L-Leu	++	++	++	++
Sauton + L-Ile	++	++	++	++
Sauton + L-Met	++	NA	++	++
Sauton + Glycine				
Group 4				
Sauton + L-Trp				++
Sauton + L-Phe				

Sauton + L-Tyr	NA
Sauton + L-Yal	NA
Sauton + L-Thr	NA

^a All amino acids, L-Ornithine and GABA supplemented to final concentration of 27 mM. NH₄Cl was tested at 1 mM, 27 mM and 96 mM.

^b Each 5 ml culture inoculated with 1x10⁷ cells of *M. smegmatis* or *M. bovis* BCG substrains.

^c Based on research literature.

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SCIENTIFIC
EDITORIAL
SERVICES

We claim:

1. A live recombinant *Mycobacterium bovis*-BCG strain comprising a nucleic acid capable of expression, the nucleic acid encoding at least one protein or polypeptide that exhibits alanine dehydrogenase activity [SEQ ID NO:1; SEQ ID NO:2], glutamine synthetase activity [SEQ ID NO:7 to SEQ ID NO:14], or L-serine dehydratase activity [SEQ ID NO:5; SEQ ID NO:6].
2. A live recombinant *Mycobacterium bovis*-BCG strain comprising a nucleic acid capable of expression, the nucleic acid encoding at least one protein or polypeptide selected from the group consisting of alanine dehydrogenase [SEQ ID NO:1; SEQ ID NO:2], glutamine synthetase [SEQ ID NO:7 to SEQ ID NO:14] and L-serine dehydratase [SEQ ID NO:5; SEQ ID NO:6].
3. A live recombinant *Mycobacterium bovis*-BCG strain comprising a nucleic acid capable of expression, the nucleic acid comprises all or part of at least one nucleic acid molecule selected from the group consisting of [SEQ ID NO:1], [SEQ ID NO:2], [SEQ ID NO:5], [SEQ ID NO:6], [SEQ ID NO:7], [SEQ ID NO:8], [SEQ ID NO:9], [SEQ ID NO:10], [SEQ ID NO:11], [SEQ ID NO:12], [SEQ ID NO:13], and [SEQ ID NO:14].
4. The live recombinant *Mycobacterium bovis*-BCG strain of claim 1, 2 or 3 wherein the *Mycobacterium bovis*-BCG strain is selected from the group consisting of *Mycobacterium bovis*-BCG-Russia, *Mycobacterium bovis*-BCG-Moreau, *Mycobacterium bovis*-BCG-Japan, *Mycobacterium bovis*-BCG-Sweden, *Mycobacterium bovis*-BCG-Birkhaug, *Mycobacterium bovis*-BCG-Prague, *Mycobacterium bovis*-BCG-Glaxo, *Mycobacterium bovis*-BCG-Denmark, *Mycobacterium bovis*-BCG-Tice, *Mycobacterium bovis*-BCG-Frappier, *Mycobacterium bovis*-BCG-Connaught, *Mycobacterium bovis*-BCG-Phipps, and *Mycobacterium bovis*-BCG-Pasteur.
5. A pharmaceutical composition comprising the live recombinant *Mycobacterium bovis*-BCG strain of claim 1, 2 or 3.

6. A vaccine or immunogenic composition for treatment or prophylaxis of a mammal against challenge by mycobacteria comprising the live recombinant *Mycobacterium bovis*-BCG strain of claim 1, 2 or 3.

7. The vaccine or immunogenic composition of claim 6 wherein the mycobacteria is *Mycobacterium tuberculosis*.

8. The vaccine or immunogenic composition of claim 6 or 7 further comprising a pharmaceutically acceptable carrier.

9. The vaccine or immunogenic composition of claim 6, 7, or 8 further comprising an adjuvant.

10. The vaccine or immunogenic composition of claim 6, 7, 8 or 9 further comprising immunogenic materials from one or more other pathogens.

11. A method for treatment or prophylaxis of a mammal against challenge by *Mycobacterium tuberculosis* or *Mycobacterium bovis* comprising administering to the mammal the vaccine or immunogenic composition of claim 1, 2 or 3.

12. The method of claim 11 wherein the mammal is a cow.

13. The method of claim 11 wherein the mammal is a human.

14. The method of claim 11 wherein the vaccine or immunogenic composition is administered in the presence of an adjuvant.

15. A method for treatment or prophylaxis of a mammal against cancer comprising administering to the mammal the vaccine or immunogenic composition of claim 1, 2 or 3.

16. The method of claim 15 wherein the vaccine or immunogenic composition is administered in the presence of an adjuvant.

17. The method of claim 15 or 16 wherein the cancer is bladder cancer.

18. A test kit comprising the live recombinant *Mycobacterium bovis*-BCG strain of claim 1, 2 or 3.

19. A media composition for inhibiting the growth of *Mycobacterium bovis*-BCG comprising alanine as the only nitrogen source for growth.

20. A media composition for inhibiting the growth of *Mycobacterium bovis*-BCG comprising serine as the only nitrogen source for growth.

21. The media composition of claim 19 or 20 further comprising:

- (a) a carbon source;
- (b) iron;
- (c) magnesium; and
- (d) SO_4 .

22. A media composition of claim 21 wherein the carbon source is selected from the group consisting of glycerol, dextrose, citrate and glucose.

23. A method for inhibiting the growth of *Mycobacterium bovis*-BCG comprising:

- (a) obtaining a sample comprising *Mycobacterium*; and
- (b) culturing the sample in a selective media.

24. The method of claim 23, wherein the selective media comprises alanine as the only nitrogen source for growth.

25. The method of claim 23, wherein the selective media comprises serine as the only nitrogen source for growth.

26. A method of culturing *Mycobacterium bovis*-BCG comprising:

- (a) obtaining a sample of *Mycobacterium*; and

(b) culturing the sample in differential media.

27. The method of claim 26, wherein the differential media comprises histidine.

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Abstract

The invention relates to a live recombinant *Mycobacterium bovis*-BCG strain comprising a nucleic acid capable of expression, the nucleic acid encoding at least one protein or polypeptide that exhibits alanine dehydrogenase activity, glutamine synthetase activity, or serine dehydratase activity.

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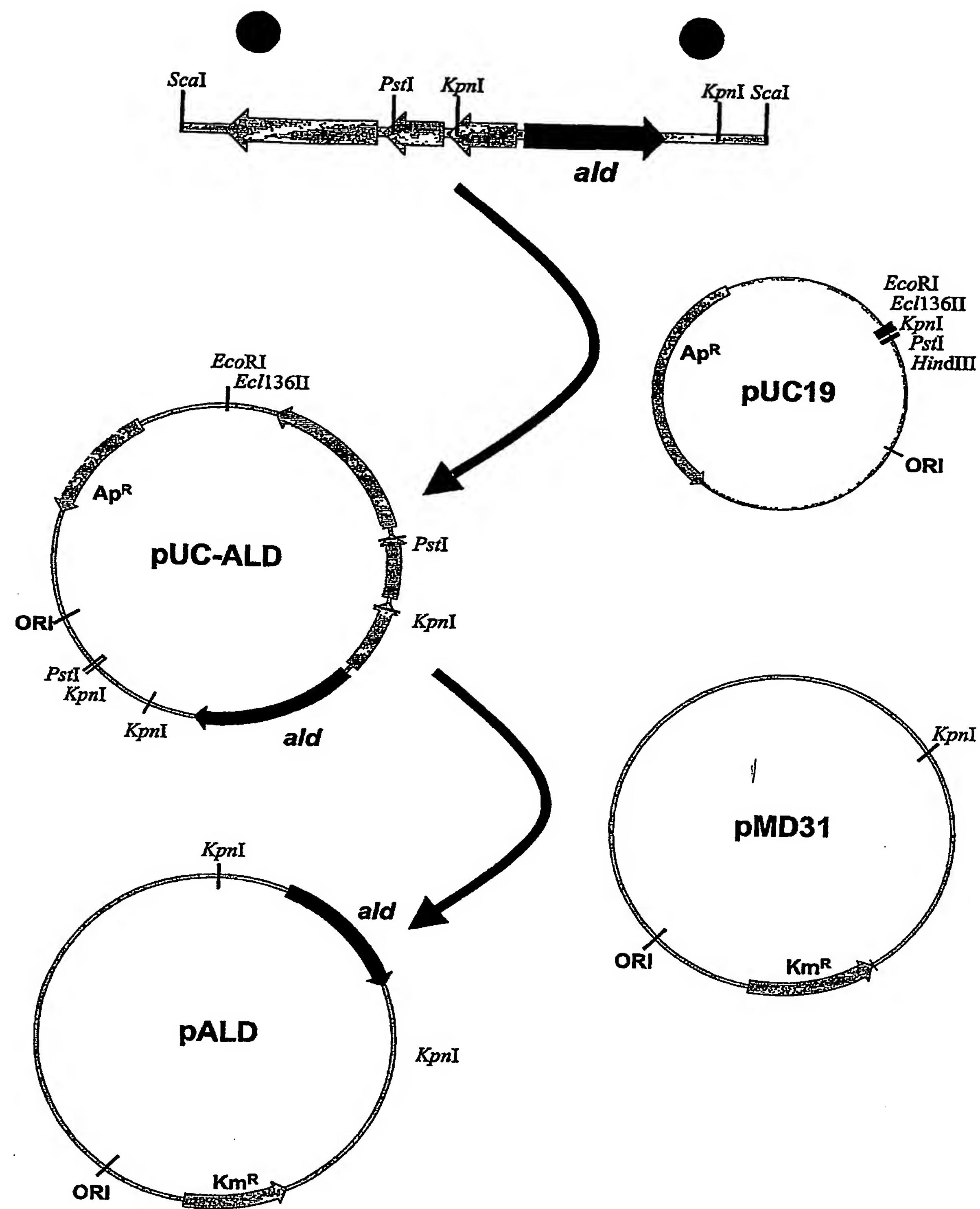


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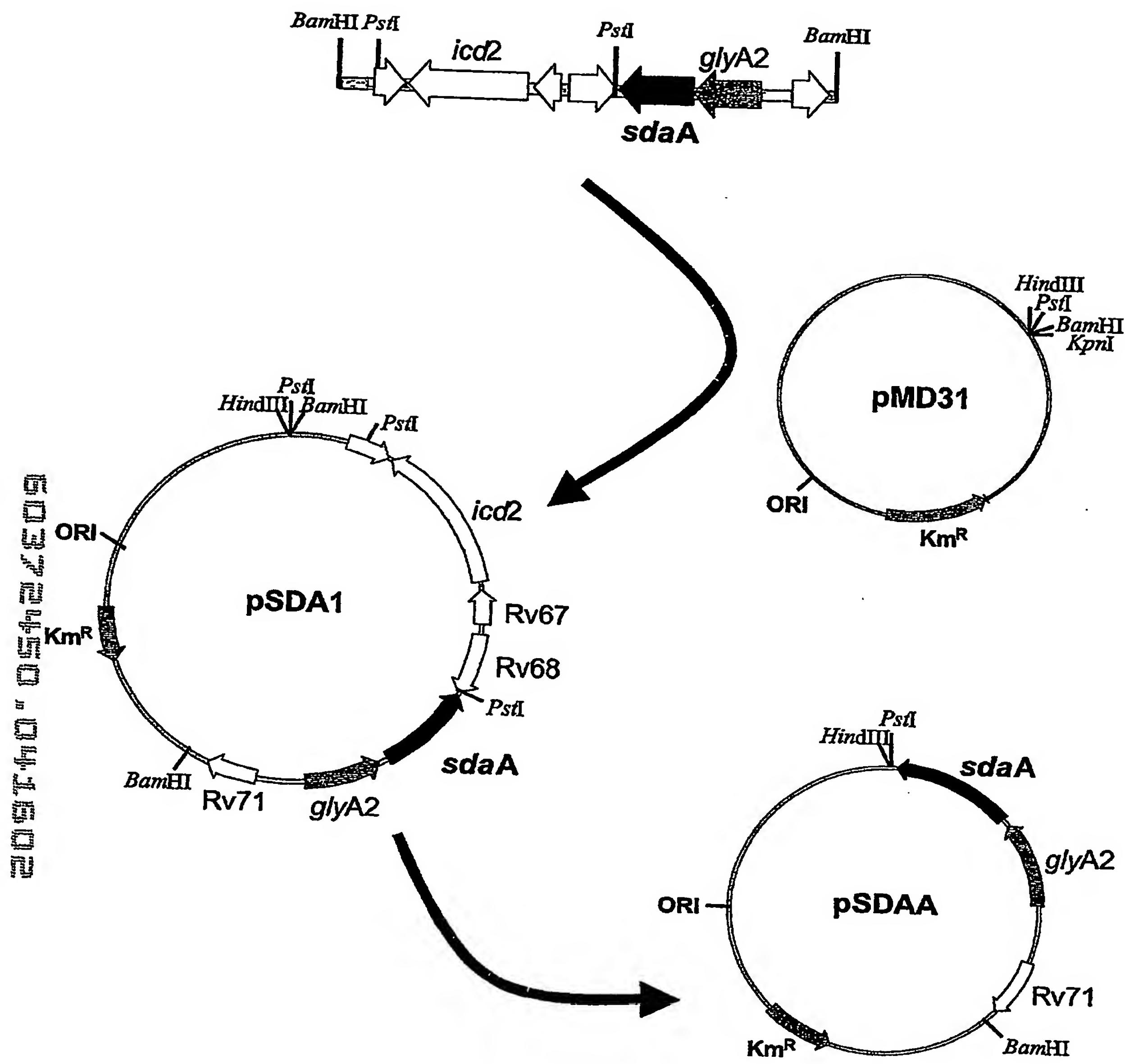


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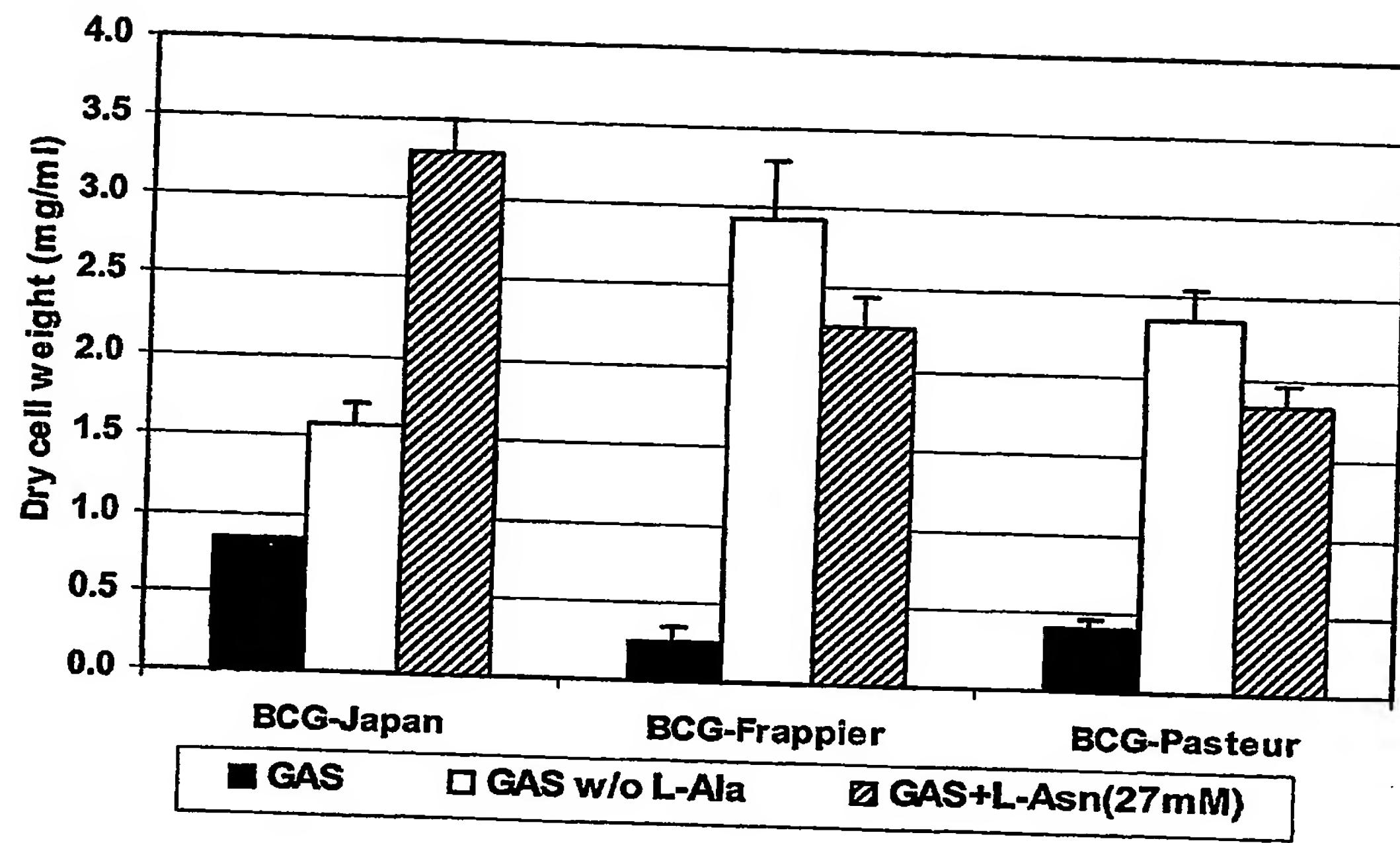


Fig. 3

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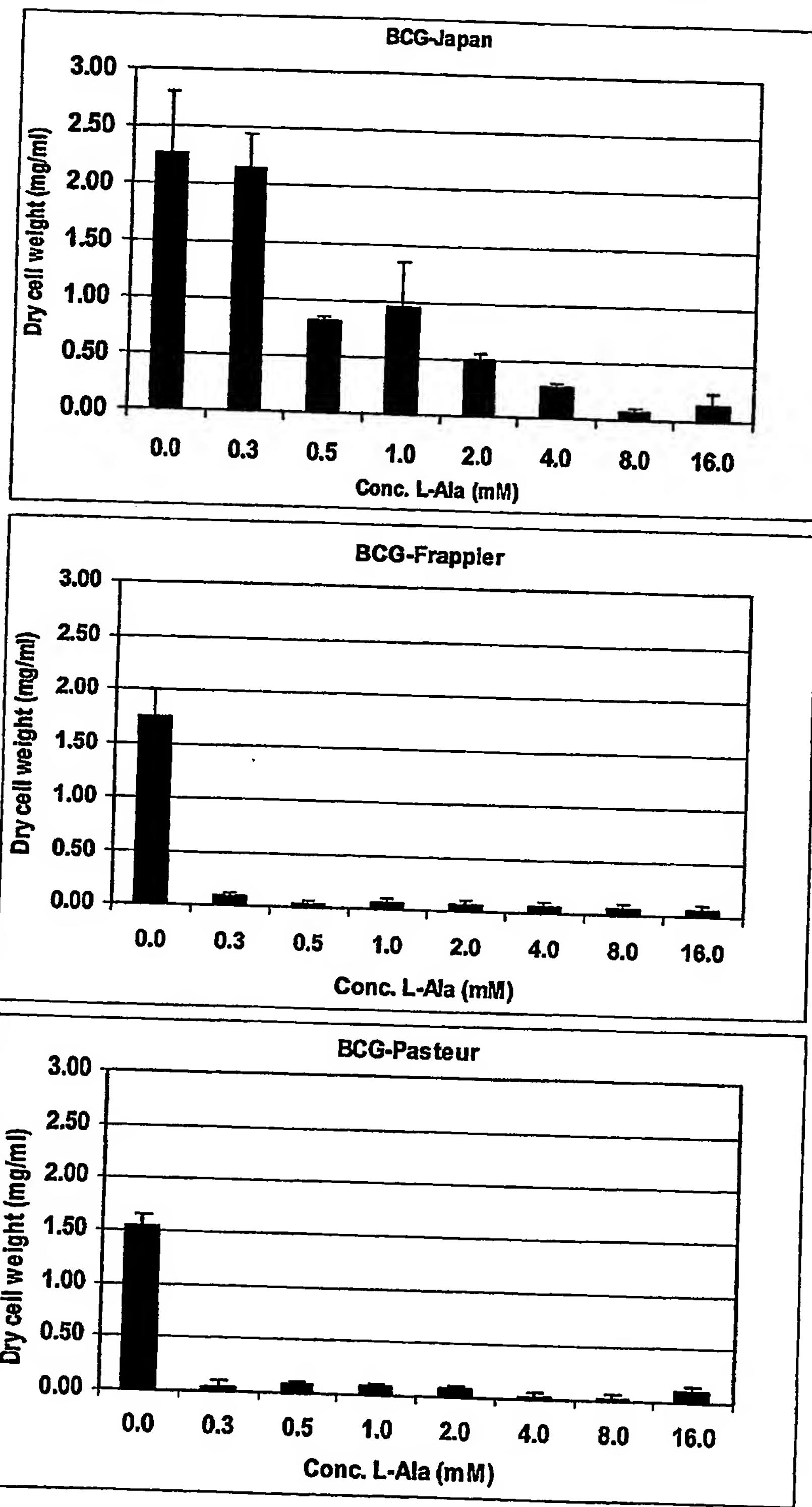


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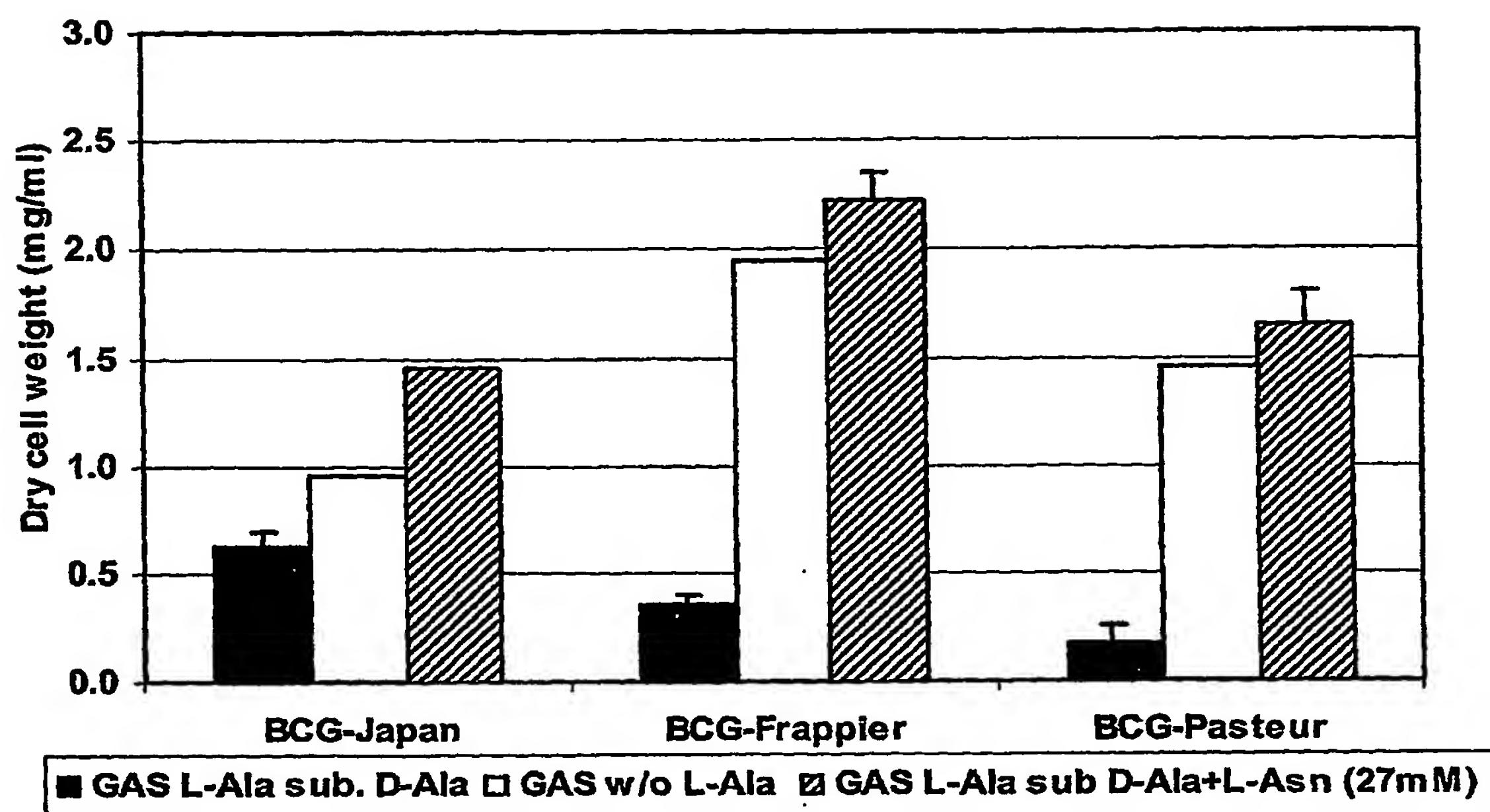


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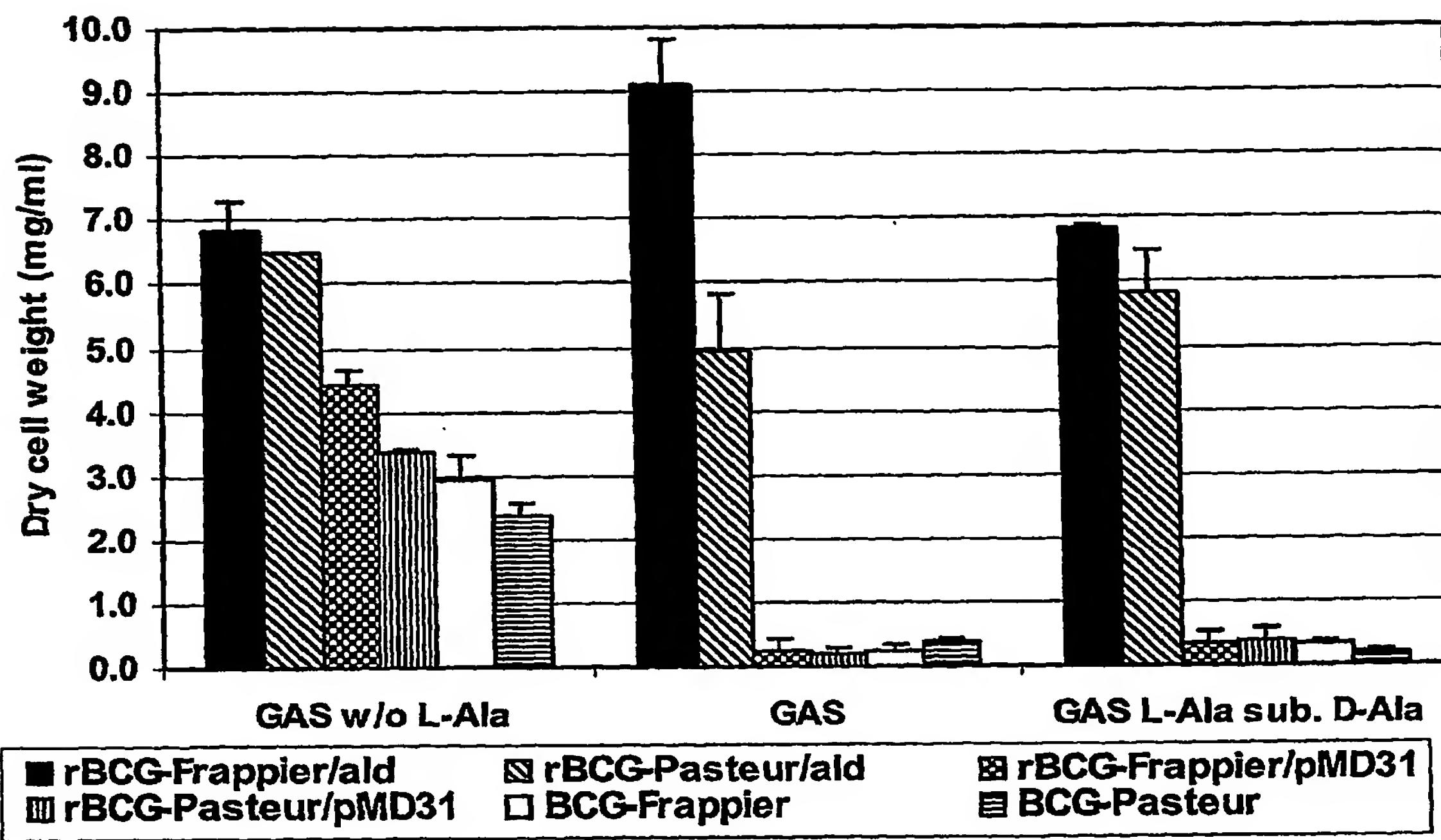


Fig. 6

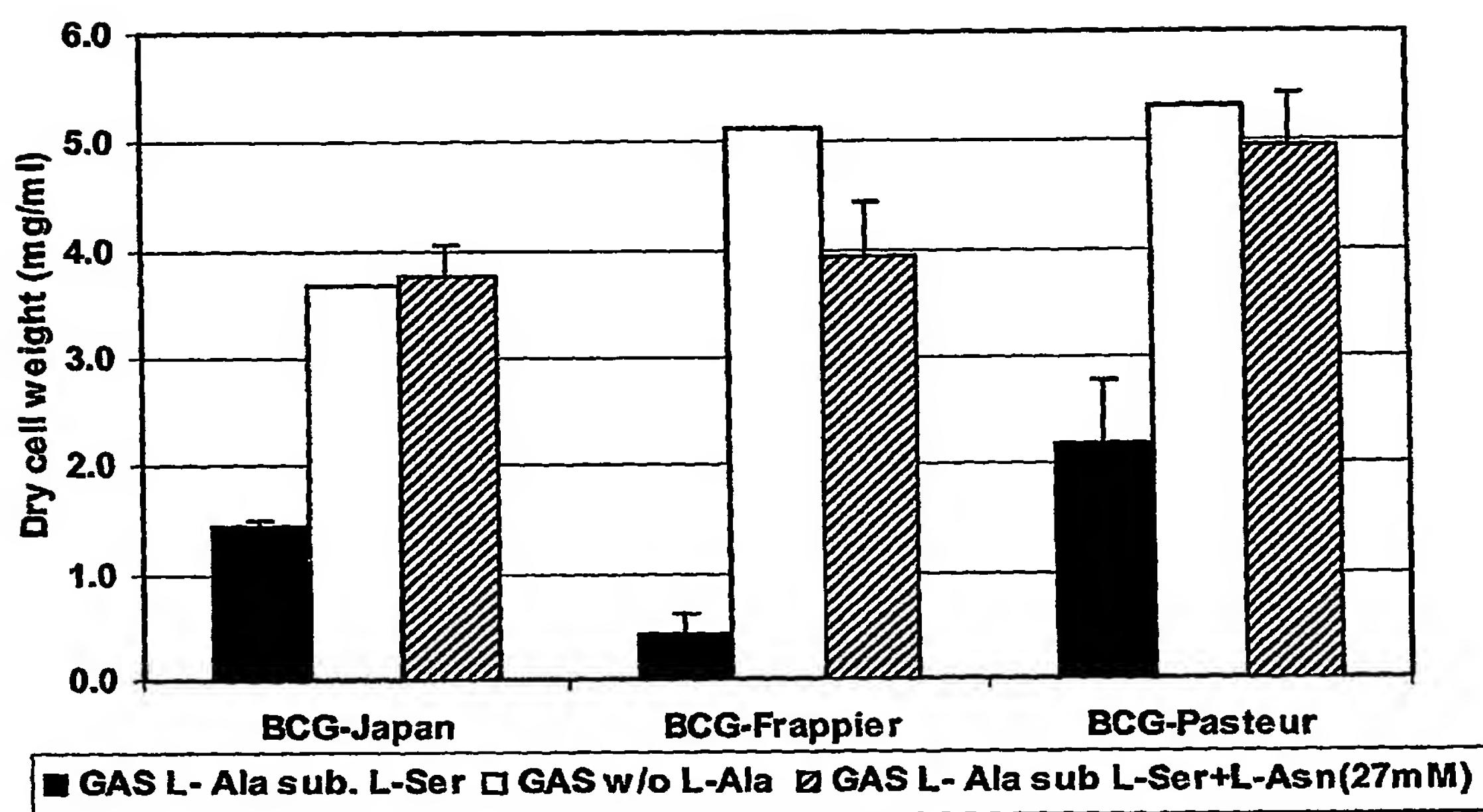


Fig. 7

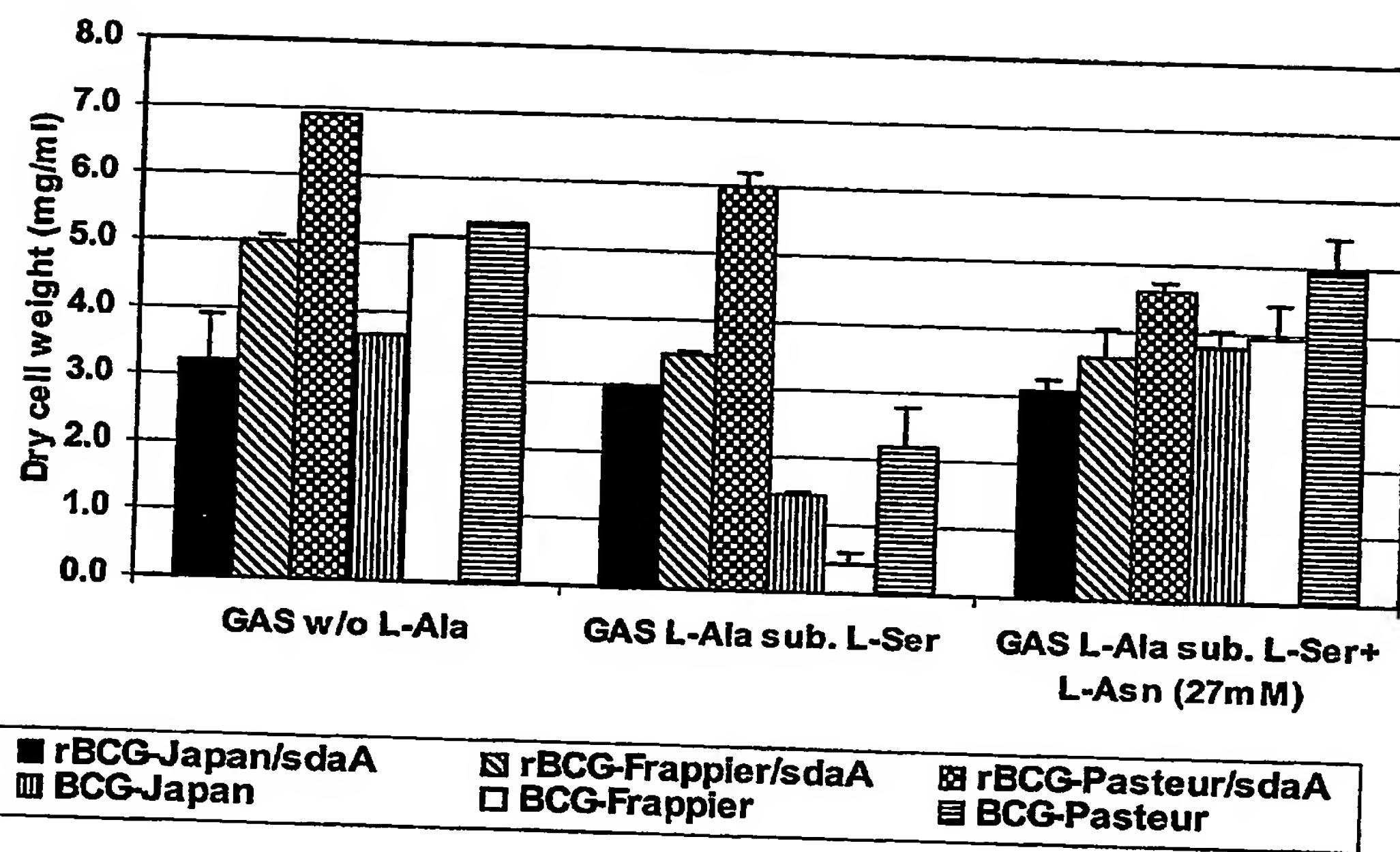


Fig. 8

A

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<i>M. bovis</i>	ATG CGC GTC GGT ATT CCG ACC GAG ACC AAA AAC AAC GAA TTC CGG GTG GCC ATC
<i>M. tb</i>	ACC CCG GCC GGC GTC GCG GAA CTA ACC CGT CGT GGC CAT GAG GTG CTC ATC CAG
<i>M. bovis</i>	ACC CCG GCC GGC GTC GCG GAA CTA ACC CGT CGT GGC CAT GAG GTG CTC ATC CAG
<i>M. tb</i>	GCA GGT GCC GGA GAG GGC TCG GCT ATC ACC GAC GCG GAT TTC AAG GCG GCA GGC
<i>M. bovis</i>	GCA GGT GCC GGA GAG GGC TCG GCT ATC ACC GAC GCG GAT TTC AAG GCG GCA GGC
<i>M. tb</i>	GCG CAA CTG GTC GGC ACC GCC GAC CAG GTG TGG GCC GAC GCT GAT TTA TTG CTC
<i>M. bovis</i>	GCG CAA CTG GTC GGC ACC GCC GAC CAG GTG TGG GCC GAC GCT GAT TTA TTG CTC
<i>M. tb</i>	AAG GTC AAA GAA CCG ATA GCG GCG GAA TAC GGC CGC CTG CGA CAC GGG CAG ATC
<i>M. bovis</i>	AAG GTC AAA GAA CCG ATA GCG GCG GAA TAC GGC CGC CTG CGA CAC GGG CAG ATC



<i>M. tb</i>	TTG TTC ACG TTC TTG CAT TTG GCC GCG TCA CGT GCT TGC ACC GAT GCG TTG TTG
<i>M. bovis</i>	TGT TCA CGT TCT TGC ATT TGG CCG CGT CAC GTG CTT GCA CCG ATG CGT TGT TGG
<i>M. tb</i>	GAT TCC GGC ACC ACG TCA ATT GCC TAC GAG ACC GTC CAG ACC GCC GAC GGC GCA
<i>M. bovis</i>	ATT CCG GCA CCA CGT CAA TTG CCT ACG AGA CCG TCC AGA CCG CCG ACG GCG CAC

<i>M. tb</i>	CTA CCC CTG CTT GCC CCG ATG AGC GAA GTC GCC GGT CGA CTC GCC GCC CAG GTT
<i>M. bovis</i>	TAC CCC TGC TTG CCC CGA TGA

<i>M. tb</i>	GGC GCT TAC CAC CTG ATG CGA ACC CAA GGG GGC CGC GGT GTG CTG ATG GGC GGG
<i>M. tb</i>	GTG CCC GGC GTC GAA CCG GCC GAC GTC GTG GTG ATC GGC GCC GGC ACC GCC GGC
<i>M. tb</i>	TAC AAC GCA GCC CGC ATC GCC AAC GGC ATG GGC GCG ACC GTT ACG GTT CTA GAC
<i>M. tb</i>	ATC AAC ATC GAC AAA CTT CGG CAA CTC GAC GCC GAG TTC TGC GCC CGG ATC CAC
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<i>M. bovis</i>	LKVKEPIAAEYGLRHGSCSRSCIWPRVLAAPRCWIPAPRQLPTRPSRPTAHPCLPR-

<i>M. tb</i>	QVGAYHLMRTQGGRGVLMGGVPGVEPADVVVIGAGTAGYNAARIANGMGTVTLDINIDKLRQLDAEFCG
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<i>M. tb</i>	ATDLGVPPTEPASVLA-
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Fig. 9

SEQUENCE LISTING

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gcc	atc	acc	ccg	gcc	ggc	gtc	g	ctg	gaa	cta	acc	cgt	cgt	ggc	cat	gag	96
Ala	Ile	Thr	Pro	Ala	Gly	Val											
																30	
gtg	ctc	atc	cag	gca	ggt	gcc	gga	gag	ggc	tcg	gct	atc	acc	gac	g		144
Val	Leu	Ile	Gln	Ala	Gly	Ala	Gly	Glu	Gly	Ser	Ala	Ile	Thr	Asp	Ala		
																45	
gat	ttc	aag	gcg	gca	ggc	g	ca	ctg	gtc	ggc	acc	gcc	gac	cag	gtg		192
Asp	Phe	Lys	Ala	Ala	Gly		Ala	Gln	Leu	Val	Gly	Thr	Ala	Asp	Gln	Val	
																60	
tgg	gcc	gac	gct	gat	tta	ttg	ctc	aag	gtc	aaa	gaa	ccg	ata	gcg	g		240
Trp	Ala	Asp	Ala	Asp	Leu	Leu	Leu	Lys	Val	Lys	Glu	Pro	Ile	Ala	Ala		
																80	
gaa	ta	ggc	cgc	ctg	cga	cac	ggg	cga	tct	tgt	tca	cgt	tct	tgc	att		288
Glu	Tyr	Gly	Arg	Leu	Arg	His	Gly	Arg	Ser	Cys	Ser	Arg	Ser	Cys	Ile		
																95	
tgg	ccg	cgt	cac	gtg	ctt	gca	ccg	atg	cgt	tgt	tgg	att	ccg	gca	cc		336
Trp	Pro	Arg	His	Val	Leu	Ala	Pro	Met	Arg	Cys	Trp	Ile	Pro	Ala	Pro		
																110	
cgt	caa	ttg	cct	acg	aga	ccg	tcc	aga	ccg	ccg	acg	gcg	cac	ta	cc		384
Arg	Gln	Leu	Pro	Thr	Arg	Pro	Ser	Arg	Pro	Pro	Pro	Thr	Ala	His	Tyr	Pro	
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tgc	ttg	ccc	cga	tga													399
Cys	Leu	Pro	Arg														
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 <212> PRT
 <213> *Mycobacterium bovis*

<400> 4

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1																15	

Ala Ile Thr Pro Ala Gly Val Ala Glu Leu Thr Arg Arg Gly His Glu
20 25 30

Val Leu Ile Gln Ala Gly Ala Gly Glu Gly Ser Ala Ile Thr Asp Ala
35 40 45

Asp Phe Lys Ala Ala Gly Ala Gln Leu Val Gly Thr Ala Asp Gln Val
50 55 60

Trp Ala Asp Ala Asp Leu Leu Leu Lys Val Lys Glu Pro Ile Ala Ala
65 70 75 80

Glu Tyr Gly Arg Leu Arg His Gly Arg Ser Cys Ser Arg Ser Cys Ile
85 90 95

Trp Pro Arg His Val Leu Ala Pro Met Arg Cys Trp Ile Pro Ala Pro
100 105 110

Arg Gln Leu Pro Thr Arg Pro Ser Arg Pro Pro Thr Ala His Tyr Pro
115 120 125

Cys Leu Pro Arg
130

<210> 5
<211> 1386
<212> DNA
<213> *Mycobacterium tuberculosis*

<220>
<221> CDS
<222> (1) .. (1386)
<223> Sequence is identical to the complement of nucleotides 13172-14551
of GenBank entry GB:MTV030 [AL021428]
Sequence is identical to the complement of nucleotides 13195-14580
of GenBank entry GB:AE006919

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Met Thr Ile Ser Val Phe Asp Leu Phe Thr Ile Gly Ile Gly Pro Ser
1 5 10 15

48

agt tcc cac acc gtg gga ccg atg cgc gcg gca aac cag ttc gta gtt
Ser Ser His Thr Val Gly Pro Met Arg Ala Ala Asn Gln Phe Val Val
20 25 30

96

gcg ctg cgc cgc cgg ggc cac ctg gat gac ctc gag gcg atg cga gtg

144

Ala Leu Arg Arg Arg Gly His Leu Asp Asp Leu Glu Ala Met Arg Val			
35	40	45	
gat ctg ttc ggc tcg ctc gcg gcc acc gga gcc ggt cat ggc acc atg			192
Asp Leu Phe Gly Ser Leu Ala Ala Thr Gly Ala Gly His Gly Thr Met			
50	55	60	
tcg gcg ata ttg ctg ggg ctg gaa ggc tgc cag cca gaa acg att acc			240
Ser Ala Ile Leu Leu Gly Leu Glu Gly Cys Gln Pro Glu Thr Ile Thr			
65	70	75	80
acc gaa cac aag gaa cgc cgg ctc gcc gag atc gca gcg tcc ggc gtg			288
Thr Glu His Lys Glu Arg Arg Leu Ala Glu Ile Ala Ala Ser Gly Val			
85	90	95	
acg cga atc ggc ggt gtc att ccg gtc ccg ctg acc gag cgt gat atc			336
Thr Arg Ile Gly Gly Val Ile Pro Val Pro Leu Thr Glu Arg Asp Ile			
100	105	110	
gac ctg cat ccc gac atc gtt ctg cca acg cat ccc aac gga atg acg			384
Asp Leu His Pro Asp Ile Val Leu Pro Thr His Pro Asn Gly Met Thr			
115	120	125	
ttc act gcc gcg ggc cca cac ggc cgc gtc ttg gcc acc gag act tat			432
Phe Thr Ala Ala Gly Pro His Gly Arg Val Leu Ala Thr Glu Thr Tyr			
130	135	140	
ttt tcg gtg ggc gga ggg ttc atc gtc acg gaa cag acc agc ggc aac			480
Phe Ser Val Gly Gly Phe Ile Val Thr Glu Gln Thr Ser Gly Asn			
145	150	155	160
agc ggc caa cat cca tgc tca gtt gcc ctt ccc tac gtg tcg gcc caa			528
Ser Gly Gln His Pro Cys Ser Val Ala Leu Pro Tyr Val Ser Ala Gln			
165	170	175	
gaa ctg ctg gac atc tgt gac cgc ctc gac gtg tca att agc gaa gcg			576
Glu Leu Leu Asp Ile Cys Asp Arg Leu Asp Val Ser Ile Ser Glu Ala			
180	185	190	
gcg ctg cgc aac gaa aca tgt tgc cgc acc gag aac gag gta cgc gcc			624
Ala Leu Arg Asn Glu Thr Cys Cys Arg Thr Glu Asn Glu Val Arg Ala			
195	200	205	
gcg ctg ctg cac ctg cgc gac gtc atg gtt gag tgc gaa cag cgg agc			672
Ala Leu Leu His Leu Arg Asp Val Met Val Glu Cys Glu Gln Arg Ser			
210	215	220	
atc gct cgc gaa ggg ttg ctt cct ggc ggc ctc cgg gtg cgc cgg cga			720
Ile Ala Arg Glu Gly Leu Leu Pro Gly Gly Leu Arg Val Arg Arg Arg			
225	230	235	240
gcg aag gtg tgg tat gac cgc ttg aac gcc gaa gac ccc act cgc aag			768
Ala Lys Val Trp Tyr Asp Arg Leu Asn Ala Glu Asp Pro Thr Arg Lys			
245	250	255	
ccg gaa ttc gct gag gac tgg gtc aac ctg gtc gcg ctg gca gtc aac			816
Pro Glu Phe Ala Glu Asp Trp Val Asn Leu Val Ala Leu Ala Val Asn			

260	265	270	
gag gag aac gcc tcc ggt ggg cgc gtc gtc acc gcc ccg acc aac ggt Glu Glu Asn Ala Ser Gly Gly Arg Val Val Thr Ala Pro Thr Asn Gly	275	280	864
		285	
gcc gcc ggc atc gtg ccg gcg gtc ctg cac tac gca atc cac tac acg Ala Ala Gly Ile Val Pro Ala Val Leu His Tyr Ala Ile His Tyr Thr	290	295	912
		300	
tcg gcc ggc gcg ggg gac ccc gac gat gtc acc gtg cga ttc ctg ctc Ser Ala Gly Ala Gly Asp Pro Asp Asp Val Thr Val Arg Phe Leu Leu	305	310	960
		315	
act gct gga gcc atc gga tcg ttg ttc aag gag cga gca tcg atc tcc Thr Ala Gly Ala Ile Gly Ser Leu Phe Lys Glu Arg Ala Ser Ile Ser	325	330	1008
		335	
gga gcc gag gtc ggc tgt cag ggc gag gtc ggc tcc gcg gcc gcc atg Gly Ala Glu Val Gly Cys Gln Gly Glu Val Gly Ser Ala Ala Ala Met	340	345	1056
		350	
gcc gcc gga ttg gct gaa atc ctc ggc ggc aca ccg cga caa gtg Ala Ala Ala Gly Leu Ala Glu Ile Leu Gly Gly Thr Pro Arg Gln Val	355	360	1104
		365	
gaa aac gcc gcc gag atc gcc atg gaa cac agc ctc ggc ctg acc tgt Glu Asn Ala Ala Glu Ile Ala Met Glu His Ser Leu Gly Leu Thr Cys	370	375	1152
		380	
gac ccc atc gcc ggg ctg gtg cag atc ccc tgc atc gaa cgc aac gcg Asp Pro Ile Ala Gly Leu Val Gln Ile Pro Cys Ile Glu Arg Asn Ala	385	390	1200
		395	
400			
att tcc gcc ggc aag gcc atc aac gcc gca cgg atg gca ttg cgc ggc Ile Ser Ala Gly Lys Ala Ile Asn Ala Ala Arg Met Ala Leu Arg Gly	405	410	1248
		415	
gac ggc atc cat cgc gtc acc ctc gac cag gtc atc gac acc atg cgc Asp Gly Ile His Arg Val Thr Leu Asp Gln Val Ile Asp Thr Met Arg	420	425	1296
		430	
gcc acc ggc gcg gac atg cac acc aag tac aag gaa acc tcg gcc ggc Ala Thr Gly Ala Asp Met His Thr Lys Tyr Lys Glu Thr Ser Ala Gly	435	440	1344
		445	
ggg ctc gcc atc aac gtc gca gtc aac atc gtc gag tgt tga Gly Leu Ala Ile Asn Val Ala Val Asn Ile Val Glu Cys	450	455	1386
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<210> 6
 <211> 461
 <212> PRT
 <213> *Mycobacterium tuberculosis*
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<221>

<222>

<223> Sequence is identical to SwissProt entry SP:SDHL_MYCTU
Sequence is identical to GenBank entries GP:AE006919_13 and GP:MTV030_11

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Ser Ser His Thr Val Gly Pro Met Arg Ala Ala Asn Gln Phe Val Val
20 25 30

Ala Leu Arg Arg Arg Gly His Leu Asp Asp Leu Glu Ala Met Arg Val
35 40 45

Asp Leu Phe Gly Ser Leu Ala Ala Thr Gly Ala Gly His Gly Thr Met
50 55 60

Ser Ala Ile Leu Leu Gly Leu Glu Gly Cys Gln Pro Glu Thr Ile Thr
65 70 75 80

Thr Glu His Lys Glu Arg Arg Leu Ala Glu Ile Ala Ala Ser Gly Val
85 90 95

Thr Arg Ile Gly Gly Val Ile Pro Val Pro Leu Thr Glu Arg Asp Ile
100 105 110

Asp Leu His Pro Asp Ile Val Leu Pro Thr His Pro Asn Gly Met Thr
115 120 125

Phe Thr Ala Ala Gly Pro His Gly Arg Val Leu Ala Thr Glu Thr Tyr
130 135 140

Phe Ser Val Gly Gly Phe Ile Val Thr Glu Gln Thr Ser Gly Asn
145 150 155 160

Ser Gly Gln His Pro Cys Ser Val Ala Leu Pro Tyr Val Ser Ala Gln
165 170 175

Glu Leu Leu Asp Ile Cys Asp Arg Leu Asp Val Ser Ile Ser Glu Ala
180 185 190

Ala Leu Arg Asn Glu Thr Cys Cys Arg Thr Glu Asn Glu Val Arg Ala
195 200 205

Ala Leu Leu His Leu Arg Asp Val Met Val Glu Cys Glu Gln Arg Ser
210 215 220

Ile Ala Arg Glu Gly Leu Leu Pro Gly Gly Leu Arg Val Arg Arg Arg
225 230 235 240

Ala Lys Val Trp Tyr Asp Arg Leu Asn Ala Glu Asp Pro Thr Arg Lys
245 250 255

Pro Glu Phe Ala Glu Asp Trp Val Asn Leu Val Ala Leu Ala Val Asn
260 265 270

Glu Glu Asn Ala Ser Gly Gly Arg Val Val Thr Ala Pro Thr Asn Gly
275 280 285

Ala Ala Gly Ile Val Pro Ala Val Leu His Tyr Ala Ile His Tyr Thr
290 295 300

Ser Ala Gly Ala Gly Asp Pro Asp Asp Val Thr Val Arg Phe Leu Leu
305 310 315 320

Thr Ala Gly Ala Ile Gly Ser Leu Phe Lys Glu Arg Ala Ser Ile Ser
325 330 335

Gly Ala Glu Val Gly Cys Gln Gly Glu Val Gly Ser Ala Ala Ala Met
340 345 350

Ala Ala Ala Gly Leu Ala Glu Ile Leu Gly Gly Thr Pro Arg Gln Val
355 360 365

Glu Asn Ala Ala Glu Ile Ala Met Glu His Ser Leu Gly Leu Thr Cys
370 375 380

Asp Pro Ile Ala Gly Leu Val Gln Ile Pro Cys Ile Glu Arg Asn Ala
385 390 395 400

Ile Ser Ala Gly Lys Ala Ile Asn Ala Ala Arg Met Ala Leu Arg Gly
405 410 415

Asp Gly Ile His Arg Val Thr Leu Asp Gln Val Ile Asp Thr Met Arg
420 425 430

Ala Thr Gly Ala Asp Met His Thr Lys Tyr Lys Glu Thr Ser Ala Gly
435 440 445

Gly Leu Ala Ile Asn Val Ala Val Asn Ile Val Glu Cys
450 455 460

<210> 7
<211> 1437
<212> DNA
<213> *Mycobacterium tuberculosis*

<220>
<221> CDS
<222> (1)...(1437)
<223> Sequence is identical to GenBank entry GB:MTU87280 [U87280]
Sequence is identical to nucleotides 163-1599 of GenBank entry GB:MTCY427
[Z70692]
Sequence is identical to nucleotides 93-1529 of GenBank entry GB:AE007073

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aag gtc gaa tat gtc gac gtc cgg ttc tgt gac ctg cct ggc atc atg		96
Lys Val Glu Tyr Val Asp Val Arg Phe Cys Asp Leu Pro Gly Ile Met		
20 25 30		
cag cac ttc acg att ccg gct tcg gcc ttt gac aag agc gtg ttt gac		144
Gln His Phe Thr Ile Pro Ala Ser Ala Phe Asp Lys Ser Val Phe Asp		
35 40 45		
gac ggc ttg gcc ttt gac ggc tcg tcg att cgc ggg ttc cag tcg atc		192
Asp Gly Leu Ala Phe Asp Gly Ser Ser Ile Arg Gly Phe Gln Ser Ile		
50 55 60		
cac gaa tcc gac atg ttg ctt ctt ccc gat ccc gag acg gcg cgc atc		240
His Glu Ser Asp Met Leu Leu Leu Pro Asp Pro Glu Thr Ala Arg Ile		
65 70 75 80		
gac ccg ttc cgc gcg gcc aag acg ctg aat atc aac ttc ttt gtg cac		288
Asp Pro Phe Arg Ala Ala Lys Thr Leu Asn Ile Asn Phe Phe Val His		
85 90 95		
gac ccg ttc acc ctg gag ccg tac tcc cgc gac ccg cgc aac atc gcc		336
Asp Pro Phe Thr Leu Glu Pro Tyr Ser Arg Asp Pro Arg Asn Ile Ala		
100 105 110		
cgc aag gcc gag aac tac ctg atc agc act ggc atc gcc gac acc gca		384
Arg Lys Ala Glu Asn Tyr Leu Ile Ser Thr Gly Ile Ala Asp Thr Ala		
115 120 125		
tac ttc ggc gcc gag gcc gag ttc tac att ttc gat tcg gtg agc ttc		432

Tyr	Phe	Gly	Ala	Glu	Ala	Glu	Phe	Tyr	Ile	Phe	Asp	Ser	Val	Ser	Phe	
130																140
gac tcg cgc gcc aac ggc tcc ttc tac gag gtg gac gcc atc tcg ggg															480	
Asp	Ser	Arg	Ala	Asn	Gly	Ser	Phe	Tyr	Glu	Val	Asp	Ala	Ile	Ser	Gly	
145																160
150																155
tgg tgg aac acc ggc gcg acc gag gcc gac ggc agt ccc aac cgg															528	
Trp	Trp	Asn	Thr	Gly	Ala	Ala	Thr	Glu	Ala	Asp	Gly	Ser	Pro	Asn	Arg	
165																175
170																
ggc tac aag gtc cgc cac aag ggc ggg tat ttc cca gtg gcc ccc aac															576	
Gly	Tyr	Lys	Val	Arg	His	Lys	Gly	Tyr	Phe	Pro	Val	Ala	Pro	Asn		
180																190
185																
gac caa tac gtc gac ctg cgc gac aag atg ctg acc aac ctg atc aac															624	
Asp	Gln	Tyr	Val	Asp	Leu	Arg	Asp	Lys	Met	Leu	Thr	Asn	Leu	Ile	Asn	
195																205
200																
tcc ggc ttc atc ctg gag aag ggc cac cac gag gtg ggc agc ggc gga															672	
Ser	Gly	Phe	Ile	Leu	Glu	Lys	Gly	His	His	Glu	Val	Gly	Ser	Gly	Gly	
210																220
215																
cag gcc gag atc aac tac cag ttc aat tcg ctg ctg cac gcc gcc gac															720	
Gln	Ala	Glu	Ile	Asn	Tyr	Gln	Phe	Asn	Ser	Leu	Leu	His	Ala	Ala	Asp	
225																240
230																
gac atg cag ttg tac aag tac atc atc aag aac acc gcc tgg cag aac															768	
Asp	Met	Gln	Leu	Tyr	Lys	Tyr	Ile	Ile	Lys	Asn	Thr	Ala	Trp	Gln	Asn	
245																255
250																
ggc aaa acg gtc acg ttc atg ccc aag ccg ctg ttc ggc gac aac ggg															816	
Gly	Lys	Thr	Val	Thr	Phe	Met	Pro	Lys	Pro	Leu	Phe	Gly	Asp	Asn	Gly	
260																270
265																
tcc ggc atg cac tgt cat cag tcg ctg tgg aag gac ggg gcc ccg ctg															864	
Ser	Gly	Met	His	Cys	His	Gln	Ser	Leu	Trp	Lys	Asp	Gly	Ala	Pro	Leu	
275																285
280																
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Met	Tyr	Asp	Glu	Thr	Gly	Tyr	Ala	Gly	Leu	Ser	Asp	Thr	Ala	Arg	His	
290																300
295																
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Tyr	Ile	Gly	Gly	Leu	Leu	His	His	Ala	Pro	Ser	Leu	Leu	Ala	Phe	Thr	
305																320
310																
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Asn	Pro	Thr	Val	Asn	Ser	Tyr	Lys	Arg	Leu	Val	Pro	Gly	Tyr	Glu	Ala	
325																335
330																
ccg atc aac ctg gtc tat agc cag cgc aac cgg tcg gca tgc gtg cgc															1056	
Pro	Ile	Asn	Leu	Val	Tyr	Ser	Gln	Arg	Asn	Arg	Ser	Ala	Cys	Val	Arg	
340																350
345																
atc ccg atc acc ggc agc aac ccg aag gcc aag cgg ctg gag ttc cga															1104	
Ile	Pro	Ile	Thr	Gly	Ser	Asn	Pro	Lys	Ala	Lys	Arg	Leu	Glu	Phe	Arg	

355	360	365	
agc ccc gac tcg tcg ggc aac ccg tat ctg gcg ttc tcg gcc atg ctg Ser Pro Asp Ser Ser Gly Asn Pro Tyr Leu Ala Phe Ser Ala Met Leu 370	375	380	1152
atg gca ggc ctg gac ggt atc aag aac aag atc gag ccg cag gcg ccc Met Ala Gly Leu Asp Gly Ile Lys Asn Lys Ile Glu Pro Gln Ala Pro 385	390	395	1200
gtc gac aag gat ctc tac gag ctg ccg gaa gag gcc gcg agt atc Val Asp Lys Asp Leu Tyr Glu Leu Pro Pro Glu Glu Ala Ala Ser Ile 405	410	415	1248
ccg cag act ccg acc cag ctg tca gat gtg atc gac cgt ctc gag gcc Pro Gln Thr Pro Thr Gln Leu Ser Asp Val Ile Asp Arg Leu Glu Ala 420	425	430	1296
gac cac gaa tac ctc acc gaa gga ggg gtg ttc aca aac gac ctg atc Asp His Glu Tyr Leu Thr Glu Gly Val Phe Thr Asn Asp Leu Ile 435	440	445	1344
gag acg tgg atc agt ttc aag cgc gaa aac gag atc gag ccg gtc aac Glu Thr Trp Ile Ser Phe Lys Arg Glu Asn Glu Ile Glu Pro Val Asn 450	455	460	1392
atc cgg ccg cat ccc tac gaa ttc gcg ctg tac tac gac gtt taa Ile Arg Pro His Pro Tyr Glu Phe Ala Leu Tyr Tyr Asp Val 465	470	475	1437
<p><210> 8</p> <p><211> 478</p> <p><212> PRT</p> <p><213> <i>Mycobacterium tuberculosis</i></p>			
<p><220></p> <p><221></p> <p><222></p> <p><223> Sequence is identical to SwissProt entry SP:GLN1_MYCTU Sequence is identical to PIR entry PIR:H70775 Sequence is identical to PRF entry PRF:2323405A</p>			
<p><400> 8</p> <p>Met Thr Glu Lys Thr Pro Asp Asp Val Phe Lys Leu Ala Lys Asp Glu 1 5 10 15</p>			
<p>Lys Val Glu Tyr Val Asp Val Arg Phe Cys Asp Leu Pro Gly Ile Met 20 25 30</p>			
<p>Gln His Phe Thr Ile Pro Ala Ser Ala Phe Asp Lys Ser Val Phe Asp 35 40 45</p>			
<p>Asp Gly Leu Ala Phe Asp Gly Ser Ser Ile Arg Gly Phe Gln Ser Ile</p>			

50

55

60

His Glu Ser Asp Met Leu Leu Leu Pro Asp Pro Glu Thr Ala Arg Ile
65 70 75 80

Asp Pro Phe Arg Ala Ala Lys Thr Leu Asn Ile Asn Phe Phe Val His
85 90 95

Asp Pro Phe Thr Leu Glu Pro Tyr Ser Arg Asp Pro Arg Asn Ile Ala
100 105 110

Arg Lys Ala Glu Asn Tyr Leu Ile Ser Thr Gly Ile Ala Asp Thr Ala
115 120 125

Tyr Phe Gly Ala Glu Ala Glu Phe Tyr Ile Phe Asp Ser Val Ser Phe
130 135 140

Asp Ser Arg Ala Asn Gly Ser Phe Tyr Glu Val Asp Ala Ile Ser Gly
145 150 155 160

Trp Trp Asn Thr Gly Ala Ala Thr Glu Ala Asp Gly Ser Pro Asn Arg
165 170 175

Gly Tyr Lys Val Arg His Lys Gly Gly Tyr Phe Pro Val Ala Pro Asn
180 185 190

Asp Gln Tyr Val Asp Leu Arg Asp Lys Met Leu Thr Asn Leu Ile Asn
195 200 205

Ser Gly Phe Ile Leu Glu Lys Gly His His Glu Val Gly Ser Gly Gly
210 215 220

Gln Ala Glu Ile Asn Tyr Gln Phe Asn Ser Leu Leu His Ala Ala Asp
225 230 235 240

Asp Met Gln Leu Tyr Lys Tyr Ile Ile Lys Asn Thr Ala Trp Gln Asn
245 250 255

Gly Lys Thr Val Thr Phe Met Pro Lys Pro Leu Phe Gly Asp Asn Gly
260 265 270

Ser Gly Met His Cys His Gln Ser Leu Trp Lys Asp Gly Ala Pro Leu
275 280 285

Met Tyr Asp Glu Thr Gly Tyr Ala Gly Leu Ser Asp Thr Ala Arg His
290 295 300

Tyr Ile Gly Gly Leu Leu His His Ala Pro Ser Leu Leu Ala Phe Thr
305 310 315 320

Asn Pro Thr Val Asn Ser Tyr Lys Arg Leu Val Pro Gly Tyr Glu Ala
325 330 335

Pro Ile Asn Leu Val Tyr Ser Gln Arg Asn Arg Ser Ala Cys Val Arg
340 345 350

Ile Pro Ile Thr Gly Ser Asn Pro Lys Ala Lys Arg Leu Glu Phe Arg
355 360 365

Ser Pro Asp Ser Ser Gly Asn Pro Tyr Leu Ala Phe Ser Ala Met Leu
370 375 380

Met Ala Gly Leu Asp Gly Ile Lys Asn Lys Ile Glu Pro Gln Ala Pro
385 390 395 400

Val Asp Lys Asp Leu Tyr Glu Leu Pro Pro Glu Glu Ala Ala Ser Ile
405 410 415

Pro Gln Thr Pro Thr Gln Leu Ser Asp Val Ile Asp Arg Leu Glu Ala
420 425 430

Asp His Glu Tyr Leu Thr Glu Gly Gly Val Phe Thr Asn Asp Leu Ile
435 440 445

Glu Thr Trp Ile Ser Phe Lys Arg Glu Asn Glu Ile Glu Pro Val Asn
450 455 460

Ile Arg Pro His Pro Tyr Glu Phe Ala Leu Tyr Tyr Asp Val
465 470 475

<210> 9
<211> 1341
<212> DNA
<213> *Mycobacterium tuberculosis*

<220>
<221> CDS

<222> (1)..(1341)

<223> Sequence is identical to complement of nucleotides 4950-6290
of GenBank entry GB:MTCY427 [Z70692]
Sequence is identical to complement of nucleotides 4880-6220
of GenBank entry GB:AE007073

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atc cgc ttc gtc cggtt cgt aca gac gtg ctc ggt ttc ctc aag	96
Ile Arg Phe Val Arg Leu Trp Phe Thr Asp Val Leu Gly Phe Leu Lys	
20 25 30	
tcg gtc gcc atc gcc cca gcc gaa ctc gag ggc gcc ttc gag gaa ggc	144
Ser Val Ala Ile Ala Pro Ala Glu Leu Glu Gly Ala Phe Glu Glu Gly	
35 40 45	
atc ggc ttc gac gga tcc tcg atc gag ggc ttt gcg cgg gtc tcg gaa	192
Ile Gly Phe Asp Gly Ser Ser Ile Glu Gly Phe Ala Arg Val Ser Glu	
50 55 60	
tcc gat acg gtg gcg cac ccg gac ccg tcg acc ttc cag gtg ctg ccc	240
Ser Asp Thr Val Ala His Pro Asp Pro Ser Thr Phe Gln Val Leu Pro	
65 70 75 80	
tgg gcc acc agt tcc ggc cac cac cac tca gcg cgg atg ttt tgc gac	288
Trp Ala Thr Ser Ser Gly His His Ser Ala Arg Met Phe Cys Asp	
85 90 95	
atc acc atg ccg gac ggc tcg ccg tcg tgg gcg gac ccg cgg cac gtg	336
Ile Thr Met Pro Asp Gly Ser Pro Ser Trp Ala Asp Pro Arg His Val	
100 105 110	
ttg cgg cgg cag ctg acg aag gcc ggc gaa ctc ggc ttc tcc tgc tac	384
Leu Arg Arg Gln Leu Thr Lys Ala Gly Glu Leu Gly Phe Ser Cys Tyr	
115 120 125	
gtg cat ccc gaa atc gag ttc ttc ctg ctc aag ccc gga ccc gag gac	432
Val His Pro Glu Ile Glu Phe Phe Leu Leu Lys Pro Gly Pro Glu Asp	
130 135 140	
ggg tcg gtg ccc gtc ccg gtc gac aac gcc ggc tat ttc gac caa gcg	480
Gly Ser Val Pro Val Pro Val Asp Asn Ala Gly Tyr Phe Asp Gln Ala	
145 150 155 160	
gtg cac gac tcc gcc ttg aac ttt cgc cgc cac gcg atc gat gcc ctg	528
Val His Asp Ser Ala Leu Asn Phe Arg Arg His Ala Ile Asp Ala Leu	
165 170 175	
gaa ttc atg ggc atc tcg gtg gag ttc agc cat cac gaa ggc gca ccc	576
Glu Phe Met Gly Ile Ser Val Glu Phe Ser His His Glu Gly Ala Pro	
180 185 190	
ggc cag cag gag atc gac ctg cgg ttt gcc gac gct ctg tog atg gct	624

Gly Gln Gln Glu Ile Asp Leu Arg Phe Ala Asp Ala Leu Ser Met Ala			
195	200	205	
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Asp Asn Val Met Thr Phe Arg Tyr Val Ile Lys Glu Val Ala Leu Glu			
210	215	220	
gag ggc gcc cgg gcg tcg ttc atg ccc aag cca ttc ggc cag cac ccg			720
Glu Gly Ala Arg Ala Ser Phe Met Pro Lys Pro Phe Gly Gln His Pro			
225	230	235	240
ggc tcg gcg atg cac acc cac atg agc ctg ttc gag ggt gat gtc aac			768
Gly Ser Ala Met His Thr His Met Ser Leu Phe Glu Gly Asp Val Asn			
245	250	255	
gcg ttc cac agc gct gat gat ccg ctg cag ctg tcg gaa gtg ggt aaa			816
Ala Phe His Ser Ala Asp Asp Pro Leu Gln Leu Ser Glu Val Gly Lys			
260	265	270	
tcg ttc atc gcc ggg atc ctg gag cac gct tgc gag atc agc gcg gtc			864
Ser Phe Ile Ala Gly Ile Leu Glu His Ala Cys Glu Ile Ser Ala Val			
275	280	285	
aca aat cag tgg gtc aac tct tac aag cgg ctg gtg cag ggc ggc gaa			912
Thr Asn Gln Trp Val Asn Ser Tyr Lys Arg Leu Val Gln Gly Gly Glu			
290	295	300	
gcg ccc acg gcc gcg tcg tgg ggg gcc aac cga tcc gcc cta gtg			960
Ala Pro Thr Ala Ala Ser Trp Gly Ala Ala Asn Arg Ser Ala Leu Val			
305	310	315	320
cgg gtg ccg atg tac acg ccg cac aag acc tcg tcg cgg cgg gtc gaa			1008
Arg Val Pro Met Tyr Thr Pro His Lys Thr Ser Ser Arg Arg Val Glu			
325	330	335	
gta cgc agc cct gat tcg gcg tgc aat ccc tat ctg aca ttc gcc gtg			1056
Val Arg Ser Pro Asp Ser Ala Cys Asn Pro Tyr Leu Thr Phe Ala Val			
340	345	350	
ctg ctg gcc gcg gga ttg cgg ggt gta gag aag ggt tac gtg ctg ggc			1104
Leu Leu Ala Ala Gly Leu Arg Gly Val Glu Lys Gly Tyr Val Leu Gly			
355	360	365	
ccg cag gcc gag gac aac gta tgg gac ctc aca ccc gag gaa cgc cga			1152
Pro Gln Ala Glu Asp Asn Val Trp Asp Leu Thr Pro Glu Glu Arg Arg			
370	375	380	
gcg atg ggg tac cga gaa ttg ccg tcc agt ttg gat agt gcg ctg cgc			1200
Ala Met Gly Tyr Arg Glu Leu Pro Ser Ser Leu Asp Ser Ala Leu Arg			
385	390	395	400
gcc atg gag gcc tcc gaa ctc gtc gcg gag gcc ttg ggg gag cac gtt			1248
Ala Met Glu Ala Ser Glu Leu Val Ala Glu Ala Leu Gly Glu His Val			
405	410	415	
ttt gac ttt ttc ttg cgc aac aag cgc acg gag tgg gcg aac tac cgc			1296
Phe Asp Phe Phe Leu Arg Asn Lys Arg Thr Glu Trp Ala Asn Tyr Arg			

420

425

430

agc cac gtc acg cca tac gag ctg cgc acc tac ctg tcg ctg tag
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435 440 445

1341

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<211> 446
<212> PRT
<213> *Mycobacterium tuberculosis*

<220>
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Sequence is identical to PIR entry PIR:B70776

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Ser Val Ala Ile Ala Pro Ala Glu Leu Glu Gly Ala Phe Glu Glu Gly
35 40 45

Ile Gly Phe Asp Gly Ser Ser Ile Glu Gly Phe Ala Arg Val Ser Glu
50 55 60

Ser Asp Thr Val Ala His Pro Asp Pro Ser Thr Phe Gln Val Leu Pro
65 70 75 80

Trp Ala Thr Ser Ser Gly His His His Ser Ala Arg Met Phe Cys Asp
85 90 95

Ile Thr Met Pro Asp Gly Ser Pro Ser Trp Ala Asp Pro Arg His Val
100 105 110

Leu Arg Arg Gln Leu Thr Lys Ala Gly Glu Leu Gly Phe Ser Cys Tyr
115 120 125

Val His Pro Glu Ile Glu Phe Phe Leu Leu Lys Pro Gly Pro Glu Asp
130 135 140

Gly Ser Val Pro Val Pro Val Asp Asn Ala Gly Tyr Phe Asp Gln Ala

145 150 155 160

Val His Asp Ser Ala Leu Asn Phe Arg Arg His Ala Ile Asp Ala Leu
165 170 175

Glu Phe Met Gly Ile Ser Val Glu Phe Ser His His Glu Gly Ala Pro
180 185 190

Gly Gln Gln Glu Ile Asp Leu Arg Phe Ala Asp Ala Leu Ser Met Ala
195 200 205

Asp Asn Val Met Thr Phe Arg Tyr Val Ile Lys Glu Val Ala Leu Glu
210 215 220

Glu Gly Ala Arg Ala Ser Phe Met Pro Lys Pro Phe Gly Gln His Pro
225 230 235 240

Gly Ser Ala Met His Thr His Met Ser Leu Phe Glu Gly Asp Val Asn
245 250 255

Ala Phe His Ser Ala Asp Asp Pro Leu Gln Leu Ser Glu Val Gly Lys
260 265 270

Ser Phe Ile Ala Gly Ile Leu Glu His Ala Cys Glu Ile Ser Ala Val
275 280 285

Thr Asn Gln Trp Val Asn Ser Tyr Lys Arg Leu Val Gln Gly Gly Glu
290 295 300

Ala Pro Thr Ala Ala Ser Trp Gly Ala Ala Asn Arg Ser Ala Leu Val
305 310 315 320

Arg Val Pro Met Tyr Thr Pro His Lys Thr Ser Ser Arg Arg Val Glu
325 330 335

Val Arg Ser Pro Asp Ser Ala Cys Asn Pro Tyr Leu Thr Phe Ala Val
340 345 350

Leu Leu Ala Ala Gly Leu Arg Gly Val Glu Lys Gly Tyr Val Leu Gly
355 360 365

Pro Gln Ala Glu Asp Asn Val Trp Asp Leu Thr Pro Glu Glu Arg Arg
370 375 380

Ala Met Gly Tyr Arg Glu Leu Pro Ser Ser Leu Asp Ser Ala Leu Arg
385 390 395 400

Ala Met Glu Ala Ser Glu Leu Val Ala Glu Ala Leu Gly Glu His Val
405 410 415

Phe Asp Phe Phe Leu Arg Asn Lys Arg Thr Glu Trp Ala Asn Tyr Arg
420 425 430

Ser His Val Thr Pro Tyr Glu Leu Arg Thr Tyr Leu Ser Leu
435 440 445

<210> 11
<211> 1353
<212> DNA
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<220>
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<222> (1)...(1353)
<223> Sequence is identical to nucleotides 4871-6223
of GenBank entry GB:MTCY180 [Z97193]
Sequence is identical to nucleotides 7308-8660
of GenBank entry GB:AE007049

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Glu Gly Val Asp Thr Val Ile Gly Thr Val Val Asn Pro Ala Gly Leu
20 25 30
acc cag gcc aag acc gtg ccg ata cgc cgg acc aac aca ttc gcc aat 144
Thr Gln Ala Lys Thr Val Pro Ile Arg Arg Thr Asn Thr Phe Ala Asn
35 40 45
cct ggc ctc ggc gcc agt ccg gtg tgg cat acc ttc tgt atc gac caa 192
Pro Gly Leu Gly Ala Ser Pro Val Trp His Thr Phe Cys Ile Asp Gln
50 55 60
tgc agt att gca ttc acc gca gac atc agt gtg gtc ggc gat caa cgt 240
Cys Ser Ile Ala Phe Thr Ala Asp Ile Ser Val Val Gly Asp Gln Arg
65 70 75 80
ctc cgc atc gat ctg tcc gcc ttg cgc atc atc ggc gac ggg ttg gcg 288
Leu Arg Ile Asp Leu Ser Ala Leu Arg Ile Ile Gly Asp Gly Leu Ala
85 90 95

tgg gcg ccc gcc ggg ttc ttc gag cag gac ggc aca ccg gtc ccc gcc Trp Ala Pro Ala Gly Phe Phe Glu Gln Asp Gly Thr Pro Val Pro Ala 100 105 110	336
tgc agc cga gga aca ctg agc cgg atc gag gcc gcg ctt gct gat gcc Cys Ser Arg Gly Thr Leu Ser Arg Ile Glu Ala Ala Leu Ala Asp Ala 115 120 125	384
ggc atc gac gcg gta atc ggc cac gaa gtc gaa ttc ctc ttg gtc gac Gly Ile Asp Ala Val Ile Gly His Glu Val Glu Phe Leu Leu Val Asp 130 135 140	432
gcg gac ggc cag cgg ctg cct tcg acg ctg tgg gcg cag tac ggt gtc Ala Asp Gly Gln Arg Leu Pro Ser Thr Leu Trp Ala Gln Tyr Gly Val 145 150 155 160	480
gcc ggg gtg ctc gag cac gag gcg ttc gtc cgc gat gtc aac gcc gcg Ala Gly Val Leu Glu His Glu Ala Phe Val Arg Asp Val Asn Ala Ala 165 170 175	528
gca acg gca gca ggc atc gct atc gag cag ttc cat ccc gaa tac ggt Ala Thr Ala Ala Gly Ile Ala Ile Glu Gln Phe His Pro Glu Tyr Gly 180 185 190	576
gcc aac caa ttc gag atc tcg tta gcg ccg cag ccg ccg gtc gcg gcc Ala Asn Gln Phe Glu Ile Ser Leu Ala Pro Gln Pro Pro Val Ala Ala 195 200 205	624
gcc gat cag ctg gtg ctg acc cgc ctc atc atc ggc cgt acc gcc cgc Ala Asp Gln Leu Val Leu Thr Arg Leu Ile Ile Gly Arg Thr Ala Arg 210 215 220	672
cgg cac ggg tta cgc gtg agc cta tcg cca gcg ccc ttc gcc gga agt Arg His Gly Leu Arg Val Ser Leu Ser Pro Ala Pro Phe Ala Gly Ser 225 230 235 240	720
atc gga tcc ggt gcc cac caa cac ttc tcg ctg act atg tcg gaa ggg Ile Gly Ser Gly Ala His Gln His Phe Ser Leu Thr Met Ser Glu Gly 245 250 255	768
atg ctg ttc tcc ggt ggg act gga gca gct ggc atg acc tcg gcc ggg Met Leu Phe Ser Gly Gly Thr Gly Ala Ala Gly Met Thr Ser Ala Gly 260 265 270	816
gag gcc gcg gtg gca gga gtg ctt cgc gga cta ccg gac gcc caa ggc Glu Ala Ala Val Ala Gly Val Leu Arg Gly Leu Pro Asp Ala Gln Gly 275 280 285	864
atc ctg tgc gga tcg atc gtg tcc ggt ctg cga atg cga ccc ggt aac Ile Leu Cys Gly Ser Ile Val Ser Gly Leu Arg Met Arg Pro Gly Asn 290 295 300	912
tgg gcc gga atc tat gca tgc tgg ggt acc gaa aac cgg gaa gcg gcg Trp Ala Gly Ile Tyr Ala Cys Trp Gly Thr Glu Asn Arg Glu Ala Ala 305 310 315 320	960
gtg cga ttc gtc aag ggc ggg gct ggc agc gcg tac ggc ggg aac gtg	1008

Val Arg Phe Val Lys Gly Gly Ala Gly Ser Ala Tyr Gly Gly Asn Val			
325	330	335	
gag gtg aag gtc gtc gac ccg tcg gcc aac ccg tat ctc gcg tcg gcg			1056
Glu Val Lys Val Val Asp Pro Ser Ala Asn Pro Tyr Leu Ala Ser Ala			
340	345	350	
gcg atc ctc gga ctg gca ctc gac ggc atg aag acc aag gcg gtg ttg			1104
Ala Ile Leu Gly Leu Ala Leu Asp Gly Met Lys Thr Lys Ala Val Leu			
355	360	365	
ccg tcg gaa acg acc gta gac ccg aca cag ctg tct gac gtg gat cgt			1152
Pro Ser Glu Thr Thr Val Asp Pro Thr Gln Leu Ser Asp Val Asp Arg			
370	375	380	
gac cgt gcc ggc att ctg cga ctt gct gcc gat cag gcg gat gca att			1200
Asp Arg Ala Gly Ile Leu Arg Leu Ala Ala Asp Gln Ala Asp Ala Ile			
385	390	395	400
gct gta ctg gat agt tcg aaa ctg ctt cgg tgc atc ctt ggc gat ccc			1248
Ala Val Leu Asp Ser Ser Lys Leu Leu Arg Cys Ile Leu Gly Asp Pro			
405	410	415	
gtg gta gat gcc gtg gtc gcg gta cgc cag tta gag cat gag cgc tac			1296
Val Val Asp Ala Val Val Ala Val Arg Gln Leu Glu His Glu Arg Tyr			
420	425	430	
ggt gac ctc gat cct gcg cag ctg gcc gac aag ttc cgg atg gct tgg			1344
Gly Asp Leu Asp Pro Ala Gln Leu Ala Asp Lys Phe Arg Met Ala Trp			
435	440	445	
agt gtg taa			1353
Ser Val			
450			
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<213> <i>Mycobacterium tuberculosis</i>			
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<222>			
<223> Sequence is identical to PIR entry PIR:C70515			
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20	25	30	
Thr Gln Ala Lys Thr Val Pro Ile Arg Arg Thr Asn Thr Phe Ala Asn			

35

40

45

Pro Gly Leu Gly Ala Ser Pro Val Trp His Thr Phe Cys Ile Asp Gln
50 55 60

Cys Ser Ile Ala Phe Thr Ala Asp Ile Ser Val Val Gly Asp Gln Arg
65 70 75 80

Leu Arg Ile Asp Leu Ser Ala Leu Arg Ile Ile Gly Asp Gly Leu Ala
85 90 95

Trp Ala Pro Ala Gly Phe Phe Glu Gln Asp Gly Thr Pro Val Pro Ala
100 105 110

Cys Ser Arg Gly Thr Leu Ser Arg Ile Glu Ala Ala Leu Ala Asp Ala
115 120 125

Gly Ile Asp Ala Val Ile Gly His Glu Val Glu Phe Leu Leu Val Asp
130 135 140

Ala Asp Gly Gln Arg Leu Pro Ser Thr Leu Trp Ala Gln Tyr Gly Val
145 150 155 160

Ala Gly Val Leu Glu His Glu Ala Phe Val Arg Asp Val Asn Ala Ala
165 170 175

Ala Thr Ala Ala Gly Ile Ala Ile Glu Gln Phe His Pro Glu Tyr Gly
180 185 190

Ala Asn Gln Phe Glu Ile Ser Leu Ala Pro Gln Pro Pro Val Ala Ala
195 200 205

Ala Asp Gln Leu Val Leu Thr Arg Leu Ile Ile Gly Arg Thr Ala Arg
210 215 220

Arg His Gly Leu Arg Val Ser Leu Ser Pro Ala Pro Phe Ala Gly Ser
225 230 235 240

Ile Gly Ser Gly Ala His Gln His Phe Ser Leu Thr Met Ser Glu Gly
245 250 255

Met Leu Phe Ser Gly Gly Thr Gly Ala Ala Gly Met Thr Ser Ala Gly
260 265 270

Glu Ala Ala Val Ala Gly Val Leu Arg Gly Leu Pro Asp Ala Gln Gly
275 280 285

Ile Leu Cys Gly Ser Ile Val Ser Gly Leu Arg Met Arg Pro Gly Asn
290 295 300

Trp Ala Gly Ile Tyr Ala Cys Trp Gly Thr Glu Asn Arg Glu Ala Ala
305 310 315 320

Val Arg Phe Val Lys Gly Gly Ala Gly Ser Ala Tyr Gly Gly Asn Val
325 330 335

Glu Val Lys Val Val Asp Pro Ser Ala Asn Pro Tyr Leu Ala Ser Ala
340 345 350

Ala Ile Leu Gly Leu Ala Leu Asp Gly Met Lys Thr Lys Ala Val Leu
355 360 365

Pro Ser Glu Thr Thr Val Asp Pro Thr Gln Leu Ser Asp Val Asp Arg
370 375 380

Asp Arg Ala Gly Ile Leu Arg Leu Ala Ala Asp Gln Ala Asp Ala Ile
385 390 395 400

Ala Val Leu Asp Ser Ser Lys Leu Leu Arg Cys Ile Leu Gly Asp Pro
405 410 415

Val Val Asp Ala Val Val Ala Val Arg Gln Leu Glu His Glu Arg Tyr
420 425 430

Gly Asp Leu Asp Pro Ala Gln Leu Ala Asp Lys Phe Arg Met Ala Trp
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Ser Val
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<210> 13
<211> 1374
<212> DNA
<213> *Mycobacterium tuberculosis*

<220>
<221> CDS

<222> (1)..(1374)

<223> Sequence is identical to complement of nucleotides 3104-4477
of GenBank entry GB:MTV003 [AL008883]
Sequence is identical to complement of nucleotides 3138-4511
of GenBank entry GB:AE007117

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ctg gtc gcg gcc ggt gac gtc gac acc gtc atc gtc gcg ttc acc gac			96
Leu Val Ala Ala Gly Asp Val Asp Thr Val Ile Val Ala Phe Thr Asp			
20 25 30			
atg cag ggc cgg ctg gcc ggc aaa cgg ata tcg ggc cgg cat ttc gtc			144
Met Gln Gly Arg Leu Ala Gly Lys Arg Ile Ser Gly Arg His Phe Val			
35 40 45			
gac gac ata gcc acc cgc ggc gtc gag tgc tgc agt tat ctg ctg gcc			192
Asp Asp Ile Ala Thr Arg Gly Val Glu Cys Cys Ser Tyr Leu Leu Ala			
50 55 60			
gtg gac gtc gac ctg aac acg gtg ccc ggc tat gcg atg gcc agt tgg			240
Val Asp Val Asp Leu Asn Thr Val Pro Gly Tyr Ala Met Ala Ser Trp			
65 70 75 80			
gac acc ggc tac ggc gat atg gtg atg acg cgg gac ttg tcc act ctg			288
Asp Thr Gly Tyr Gly Asp Met Val Met Thr Pro Asp Leu Ser Thr Leu			
85 90 95			
cgg ctg att cct tgg cta ccg gga acg gcg ctg gtg atc gcc gac ctg			336
Arg Leu Ile Pro Trp Leu Pro Gly Thr Ala Leu Val Ile Ala Asp Leu			
100 105 110			
gtc tgg gcc gac ggc agc gag gtc gcc gtc tgg ccg cgc agc att ctg			384
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115 120 125			
cgc cgt cag ctc gat cgg ctc aag gcg cgc gga ctg gtc gcc gat gtg			432
Arg Arg Gln Leu Asp Arg Leu Lys Ala Arg Gly Leu Val Ala Asp Val			
130 135 140			
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Ala Thr Glu Leu Glu Phe Ile Val Phe Asp Gln Pro Tyr Arg Gln Ala			
145 150 155 160			
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Trp Ala Ser Gly Tyr Arg Gly Leu Thr Pro Ala Ser Asp Tyr Asn Ile			
165 170 175			
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180 185 190			
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Ile Arg Leu Gly Met Ala Gly Ala Gly Leu Arg Phe Glu Ala Val Lys		
195	200	205
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Gly Glu Cys Asn Met Gly Gln Gln Glu Ile Gly Phe Arg Tyr Asp Glu		
210	215	220
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225	230	235
gaa atc gcc gac cag cac ggc aag agc cta acg ttc atg gcg aaa tac		768
Glu Ile Ala Asp Gln His Gly Lys Ser Leu Thr Phe Met Ala Lys Tyr		
245	250	255
gat gaa cgc gaa ggt aat agc tgt cac atc cat gtc tcg ctg cgt ggc		816
Asp Glu Arg Glu Gly Asn Ser Cys His Ile His Val Ser Leu Arg Gly		
260	265	270
acg gat ggc tcc gcg gtg ttt gcc gac agt aac ggg ccg cac ggc atg		864
Thr Asp Gly Ser Ala Val Phe Ala Asp Ser Asn Gly Pro His Gly Met		
275	280	285
tcg tcg atg ttc cgc agc ttc gtc gcc ggc cag ttg gcc acg ttg cgc		912
Ser Ser Met Phe Arg Ser Phe Val Ala Gly Gln Leu Ala Thr Leu Arg		
290	295	300
gaa ttc acg ctg tgc tat gcg ccg acc att aac tcc tac aag cga ttt		960
Glu Phe Thr Leu Cys Tyr Ala Pro Thr Ile Asn Ser Tyr Lys Arg Phe		
305	310	315
320		
gcc gat agc agt ttc gcg ccg acg gcg ctg gct tgg ggg ctg gac aat		1008
Ala Asp Ser Ser Phe Ala Pro Thr Ala Leu Ala Trp Gly Leu Asp Asn		
325	330	335
cgc acc tgc gcc ctg cgg gtg gtt ggc cac ggg caa aac atc cgg gtc		1056
Arg Thr Cys Ala Leu Arg Val Val Gly His Gly Gln Asn Ile Arg Val		
340	345	350
gaa tgc cgg gtt ccc ggc ggt gat gtc aac cag tac ctg gcg gtg gcg		1104
Glu Cys Arg Val Pro Gly Gly Asp Val Asn Gln Tyr Leu Ala Val Ala		
355	360	365
gct ctc att gct gga ggg ttg tac ggt atc gag cgg ggc ctt cag ctg		1152
Ala Leu Ile Ala Gly Gly Leu Tyr Gly Ile Glu Arg Gly Leu Gln Leu		
370	375	380
ccc gag ccc tgt gtc ggc aac gcc tac caa ggc gcc gat gtc gaa cgg		1200
Pro Glu Pro Cys Val Gly Asn Ala Tyr Gln Gly Ala Asp Val Glu Arg		
385	390	395
400		
ctg ccg gtt acg ctg gcc gac gcc gcg gtg ctg ttc gag gat tct gcg		1248
Leu Pro Val Thr Leu Ala Asp Ala Ala Val Leu Phe Glu Asp Ser Ala		
405	410	415
ctg gtg cgc gag gcg ttc ggc gag gat gtt gtc gcg cac tac ctg aac		1296
Leu Val Arg Glu Ala Phe Gly Glu Asp Val Val Ala His Tyr Leu Asn		

420

425

430

aac gcg cgt gtg gag ctg gcg gcg ttc aac gcg gcg gtc acc gat tgg
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1344

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450 455

1374

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<212> PRT
<213> *Mycobacterium tuberculosis*

<220>
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35 40 45

Asp Asp Ile Ala Thr Arg Gly Val Glu Cys Cys Ser Tyr Leu Leu Ala
50 55 60

Val Asp Val Asp Leu Asn Thr Val Pro Gly Tyr Ala Met Ala Ser Trp
65 70 75 80

Asp Thr Gly Tyr Gly Asp Met Val Met Thr Pro Asp Leu Ser Thr Leu
85 90 95

Arg Leu Ile Pro Trp Leu Pro Gly Thr Ala Leu Val Ile Ala Asp Leu
100 105 110

Val Trp Ala Asp Gly Ser Glu Val Ala Val Ser Pro Arg Ser Ile Leu
115 120 125

Arg Arg Gln Leu Asp Arg Leu Lys Ala Arg Gly Leu Val Ala Asp Val
130 135 140

Ala Thr Glu Leu Glu Phe Ile Val Phe Asp Gln Pro Tyr Arg Gln Ala
145 150 155 160

Trp Ala Ser Gly Tyr Arg Gly Leu Thr Pro Ala Ser Asp Tyr Asn Ile
165 170 175

Asp Tyr Ala Ile Leu Ala Ser Ser Arg Met Glu Pro Leu Leu Arg Asp
180 185 190

Ile Arg Leu Gly Met Ala Gly Ala Gly Leu Arg Phe Glu Ala Val Lys
195 200 205

Gly Glu Cys Asn Met Gly Gln Gln Glu Ile Gly Phe Arg Tyr Asp Glu
210 215 220

Ala Leu Val Thr Cys Asp Asn His Ala Ile Tyr Lys Asn Gly Ala Lys
225 230 235 240

Glu Ile Ala Asp Gln His Gly Lys Ser Leu Thr Phe Met Ala Lys Tyr
245 250 255

Asp Glu Arg Glu Gly Asn Ser Cys His Ile His Val Ser Leu Arg Gly
260 265 270

Thr Asp Gly Ser Ala Val Phe Ala Asp Ser Asn Gly Pro His Gly Met
275 280 285

Ser Ser Met Phe Arg Ser Phe Val Ala Gly Gln Leu Ala Thr Leu Arg
290 295 300

Glu Phe Thr Leu Cys Tyr Ala Pro Thr Ile Asn Ser Tyr Lys Arg Phe
305 310 315 320

Ala Asp Ser Ser Phe Ala Pro Thr Ala Leu Ala Trp Gly Leu Asp Asn
325 330 335

Arg Thr Cys Ala Leu Arg Val Val Gly His Gly Gln Asn Ile Arg Val
340 345 350

Glu Cys Arg Val Pro Gly Gly Asp Val Asn Gln Tyr Leu Ala Val Ala
355 360 365

Ala Leu Ile Ala Gly Gly Leu Tyr Gly Ile Glu Arg Gly Leu Gln Leu
370 375 380

Pro Glu Pro Cys Val Gly Asn Ala Tyr Gln Gly Ala Asp Val Glu Arg
385 390 395 400

Leu Pro Val Thr Leu Ala Asp Ala Ala Val Leu Phe Glu Asp Ser Ala
405 410 415

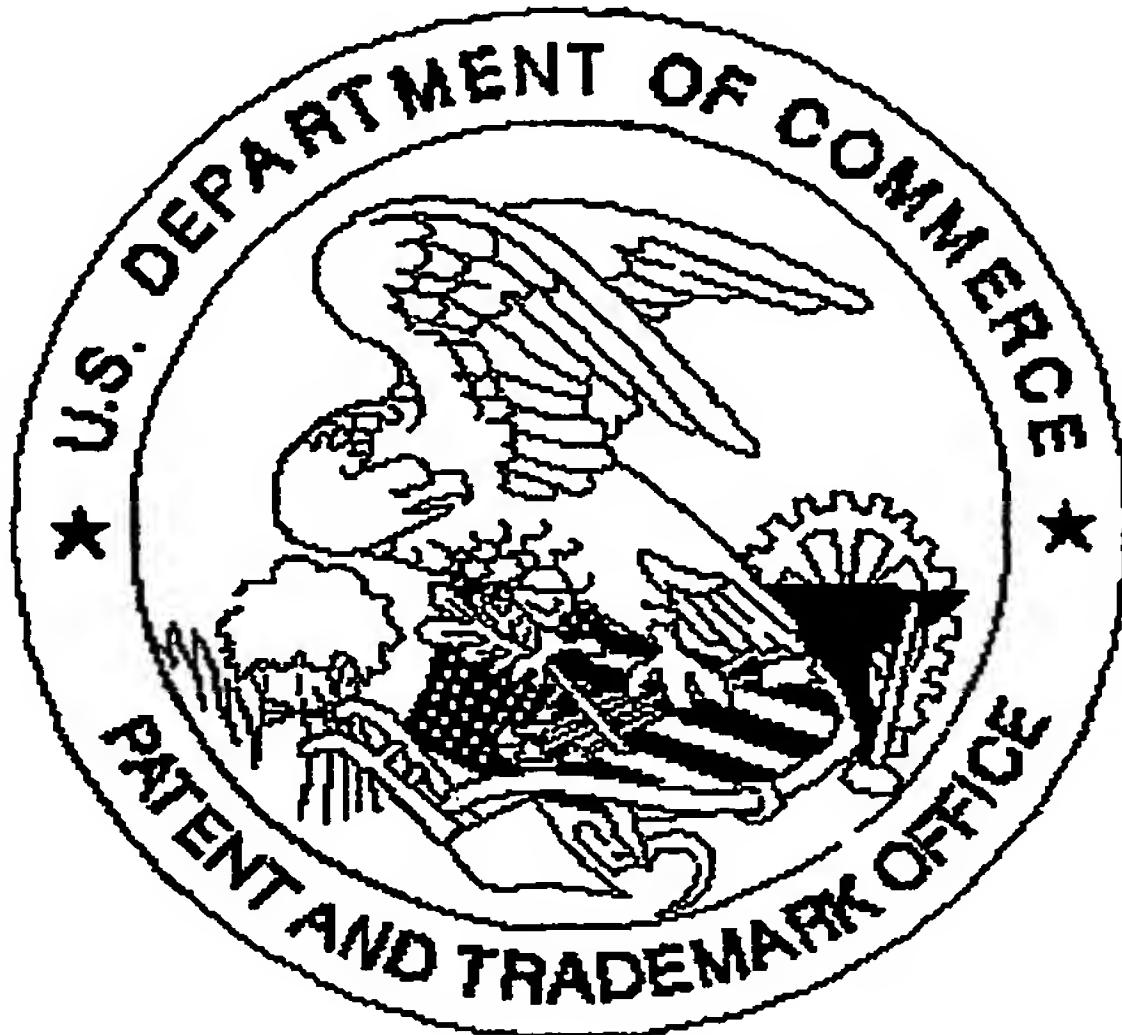
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420 425 430

Asn Ala Arg Val Glu Leu Ala Ala Phe Asn Ala Ala Val Thr Asp Trp
435 440 445

Glu Arg Ile Arg Gly Phe Glu Arg Leu
450 455

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